

# **MASS REARING OF SELECTED BUTTERFLIES FOR POSSIBLE REINTRODUCTION IN CONSERVATION PROGRAMMES**

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## **ABSTRACT**

**C**aptive breeding of organisms for possible reintroduction in species conservation programmes is well recognized. Insect breeding Centers, particularly butterfly farms, are the major attraction in many countries. However; in India not much progress has been made on these lines. In the present study, an attempt was made to standardise a methodology for captive breeding of butterflies under Kerala conditions.

General information on the butterflies found in a given locality, their habitat preferences as well as factors affecting survival is very essential for the success of any captive breeding programme. Build up of infra-structure and standardisation of a breeding programme for the species selected are also very important.

In this study, a general survey was made on species found in KFRI campus at different seasons and their number monitored. Based on data gathered, the common rose (*Pachliopta aristolochiae*) and the southern birdwing (*Troides minos*) which breed on *Aristolochia indica* were selected for mass rearing. Attempts to maintain their population throughout the year in the insecto was not successful mainly due to defects of the insecto and therefore attempts were made to lure butterflies into the garden by planting appropriate larval and adult host plants and by offering suitable ecological conditions through landscaping. General facilities required for a 'butterflyhouse' are also briefly discussed.

**Key words:** Butterfly house, butterfly garden, conservation, Kerala. India.

## INTRODUCTION

Domestication of both plant and animal species has been practised in many fields of human endeavor such as commerce, research and teaching. The range of domesticated species has increased considerably with the advancement of scientific knowledge. The classical examples of successful domestication are those of the silk worm *Bombyx mori*, the honey bee *Apis mellifera* and the medicinal leech *Hirudo medicinalis* (Wilson, 1971). Other species such as the fruit fly *Drosophila melanogaster*, locusts and the giant snail *Achatina* sp., have been bred for educational or research purposes.

Current estimates indicate that only 1.6 million out of an estimated total of 30 million species have been so far collected and identified. Of these, the invertebrates form an overwhelming majority of life on earth, in terms of individuals, species and biomass. Not only are invertebrates dominantly diverse but they are also the basis of most ecosystem processes. It has been stated that if the vertebrates were to be wiped out, the World's ecosystems would be perturbed for several years but if invertebrates were to be wiped out our planet would never recover (Wilson, 1987).

Among invertebrates, insects form the major group. According to an estimate 7,51,000 species of insects have been so far recorded (WRI, *et al*, 1992). Insects are of diversified habits and may be phytophagous, predatory, parasitic or saprophytic. However, majority of insects are phytophagous feeding either on the foliage, stem or fruits. It has been stated that approximately one quarter of insects are plant feeding species (Strong *et al* 1984). Roughly half of these belong to the group Lepidoptera which include the butterflies and the moths.

### SIGNIFICANCE OF LEPIDOPTERA

Lepidopterous insects being highly colorful have great aesthetic value. Many species of butterflies are much sought after because of their marvellous appearance. They have also been looked upon as organisms of ecological significance since they form the primary herbivores in the ecosystem. All stages of Lepidoptera are also fed upon by various higher groups of animals like birds, bats and mammals and thus form more than one link in the food web. Butterflies and moths being important pollinators of several wild and domesticated plant species, depletion of their population could adversely affect the regeneration of plants they pollinate. As the immature stages of

Lepidoptera are phytophagous in habits they also have great economic importance as pests of various crops of agricultural and forestry importance.

## **THREATS FOR SURVIVAL OF BUTTERFLIES**

### **Ecological factors**

Since butterflies are often dependent on specific host plants and have a complex life cycle, they are vulnerable to the activities of man which disturb their habitat. Land, drainage, application of fertilizers and herbicides, and mechanization have helped farmers to achieve high productivity, but these activities have affected the survival of several butterflies and their biotopes (van der Made, 1987). The Homerus Swallowtail *Papilio homerus* Fb. which once inhabited the 13 parishes in the island of Jamaica is currently found only in two isolated and diminishing strongholds. The principal reason for the depletion of this butterfly is due to destruction of the virgin wet rainforest habitat (Emmel and Garraway, 1990). Thus the main causes of the decline of butterflies include biotope destruction due to changed land management and the failure by several species to track the patches of their habitat that are still being generated in modern fragmented landscapes.

Local extinctions of butterfly populations during periods of unfavourable weather have also been reported. During such periods the habitat suitability also declines affecting survival of butterfly populations (Pollard, 1979; Thomas, 1980). For instance, the extinction of the black hairstreak *Strymonidia pruni* is reported to be caused by destruction of its breeding sites. There are about 12,000 species of butterflies found all over the world and if we assume that one in ten could be categorized as threatened, then we are obliged to suspect that more than 1,200 species of butterflies could be facing extinction. It has been estimated that extinction of a single species may set in motion a chain of events, adversely affecting the survival of 10-20 other species which are related to it.

### **Butterfly trade**

Butterflies have been objects for trade and it has been pointed out that collecting on a commercial scale can cause extinction of species (Sheldon, 1925; McLeod, 1979). Ford (1945) has stated that the extinction of *Lycaena dispar* in Britain was due to over collecting. Some are of the opinion that collecting can affect only populations that are already weakened (Ner, 1974; Spooner, 1963). Since all collections are aimed at the rare and colourful species, these are the species facing maximum threat.

## **PROTECTION OF BUTTERFLIES**

### **Through legislation**

In order to prevent over exploitation of various species, many countries have formulated strict measures of protection of fauna through legislation. Under the Indian Wildlife Act (GOI. 1982), 129 species have been included in schedule I and 292 species under schedule II. The act prohibits export of butterflies (dead or alive) and manufacture of decoration articles using these insects. According to section 40(2) of this act no person can acquire, receive, keep in custody or possession any of the species included in the above schedule without prior permission of the Chief Wildlife Warden. Although attempts are being made to prevent illicit collection of butterflies through legislation, many cases go unnoticed.

### **Through habitat improvement and reintroduction**

As has been stated earlier, habitat loss is another reason for the dwindling butterfly population. Creation of lost habitats for augmenting natural populations and reintroduction of species has been suggested for conservation of butterflies. However, this method has rarely been explored as a means to re-establish wild populations on account of the prohibitive costs (Hughes and Bennet, 1991).

Successful reintroduction programme requires establishment of appropriate technology and also formulation of a long term conservation programme for rare and endangered species. This is possible only through the establishment of specific Centers of Research and Development. At present, insect farming has been practised successfully for a few species like honey bees and silk moths and there are several research Institutions in India to cater to the R & D requirements of farming these insects. Butterfly farming Centers are not yet established in India, although in several countries like Malaysia, Taiwan, Papua New Guinea, Philippines and Sri Lanka such Centers are playing a major role in the conservation of local fauna. Such Centers may also be useful to impart environmental education to children and public (Khoshoo,

Although 269 international programmes are currently involved with species conservation, only one is concerned with invertebrates (Hughes and Bennett, 1991). It has been estimated that 2250 invertebrates (compared with 689 mammals, 1047 birds, 191 reptiles and 63 amphibians) are in danger of extinction (IUCN, 1990). With a view to promote conservation of invertebrates

IUCN has formed various 'specialists groups' which undertake captive breeding programmes. In India, very few attempts have been made in this direction except for the recent initiative by the Zoo Outreach Organization in Coimbatore.

## **SCOPE OF BUTTERFLY CONSERVATION IN INDIA**

India, with its diversified habitats is very rich in its butterfly fauna with over 1430 species of butterflies reported from all over the country. But the advancements in human civilization has affected the natural habitats of many species of butterflies threatening their survival. As a result, the range of many species are very much restricted. According to Talbot (1939, 1947) and Wynter - Blyth (1957), as much as 520 species (including subspecies and races) constituting over 30% of known butterflies are rare. Even in the natural habitats, many species having highly specific ecological requirements are facing threats due to slow changes in the ecosystem as revealed by a study in the Silent Valley National Park where of the 100 species observed, about 13 were extremely rare (Mathew, 1990).

## REVIEW OF LITERATURE

### BUTTERFLY FAUNA OF INDIA

**T**he Order Lepidoptera which is the second largest insect Order is estimated to contain over 1.40,000 species of which 12,000 are butterflies. Out of these, about 1430 species of butterflies under 10 families have been recorded from the Indian sub region (Talbot. 1939, 1947, D'Abbrera, 1982, '85, '86). In Kerala, about 320 species of butterflies have so far been reported. Due to destruction of natural habitats, a number of these butterflies are to be found only in the forests. A recent study in Nilgiri Biosphere Reserve has indicated survival of several endangered and rare species of butterflies in this region (ZSI, 1988; Larsen, 1987, 1988; Mathew, 1990). With the current rate of disturbance to natural ecosystems, many species are likely to be phased out by the turn of the century unless urgent and active remedial action is initiated. Brief account of the various families well represented in Kerala are given below:

#### **Papilionidae**

They are popularly known as 'swallow tails' on account of the tail-like extension of their hind wings. In size, they range from small to very large and are beautifully colored. Of about 217 species (including sub species), 18 species are commonly found in Kerala. The larvae of some papilionids are known to develop on plants belonging to Aristolochiaceae (*Aristolochia indica*, *Bragantia* sp.); Anonaceae (*Anona* sp., *Polyalthia* sp.); Lauraceae (*Machilus macarantha*, *Listea floribunda*, *Cinnamomum zeylanicum*, *Actinodaphne* sp.) and Rutaceae (*Citrus* sp., *Aegle marmelos*, *Atlantia* sp., *Glycosmis pentaphylla* and *Xanthoxylum rhetsa*).

The southern bird wing (*Troidesminos*), the malabar rose (*Pachliopta jophon*), the common rose (*P. aristolochiae*), the common mime (*Chilasa clytia*), the blue mormon (*Papilio polymnestor*) are some common papilionid butterflies found in Kerala.

#### **Pieridae**

Commonly known as whites or yellows, most of them are fond of sunshine and are found to frequent banks of streams or puddles in aggregations. Of the 100 species reported from India, 35 species are found in Kerala.



The larvae develop on plants belonging to the families, Berberidaceae (*Berberis* sp.). Capparidaceae (*Capparis* sp., *Crataeva* sp., *Cadaba* sp. and *Maerua* sp.); Cruciferae (*Cabbage*); Euphorbiaceae (*Hemicyclia* sp.) Leguminosae (*Cassia* sp., *Albizia* sp., *Wagatea* sp., *Poinciana* sp., *Butea* sp.); Loranthaceae (*Loranthus*) as well as Salvadoraceae (*Salvadora* sp.).

The common jezebel (*Delias eucharis*), the common wanderer (*Pareronia valeria*), the lemon emigrant (*Catopsilia pyranthe*) and the common emigrant (*Catopsilia pomona*), the mottled emigrant (*Catopsilia pyranthe*) and the common grass yellow (*Eurema hecabe*) are the common pierids found in Kerala.

## **Danaidae**

These are relatively hard butterflies that are unpalatable to predators on account of their feeding on plants containing alkaloids. The caterpillars develop mostly on plants belonging to Apocyanaceae (*Agonosma* sp., *Vallisneria* sp., *Ichocarpis* sp., *Marsdenia* sp. and *Raphistemma* sp.); Rubiaceae (*Allophania* sp.); Asclepiadaceae, Caesalpinaceae and Utricaceae (*Ficus* spp.).

There are 77 species occurring in the Indian subcontinent of which 14 species are found in Kerala. The Malabar tree nymph (*Idea malabarica*), the nilgiri tiger (*Parantica nilgiriensis*), the blue tiger (*P. limniace*), the glassy blue tiger (*P. aglea*), the striped tiger (*Danus genivita*), the plain tiger (*D. chrysippus*) and the common crow (*Euploeacore*) are certain species found in Kerala.

## **Satyridae**

These butterflies are predominantly shade loving species and are found mostly in dense forests or such situations. They are mostly brownish in color and occur in large numbers frequenting over-ripe fruits or sappy exudation of plants. Some species exhibit seasonal variations as dry and wet season forms. Of about 330 species reported from the Indian subcontinent, 30 are found in Kerala which include the common bush brown (*Mycalesis perseus*), the white bar bush brown (*M. anaxias*), the common tree brown (*Lethe rohria*) and the common five ring (*Ypthima baldus*). Larvae develop on palms, grasses and bamboos.

## **Amathusidae**

Members of this family, generally known as jungle queen, palm kings, duffers etc., are large sized and sombre colored. The morpho butterflies of South

America are well known for their brilliant iridescent colors. They rarely venture out in bright sunlight and are often confined to thick forests. Adults feed on over-ripe fruits and sappy exudation of trees. Their immature forms feed on leaves of bamboos or palms.

The species found in the Indo-Malayan region are very colorful and there are about 50 species in the Indian sub region of which at least two species belonging to Discophora are common in Kerala.

## **Nymphalidae**

These butterflies are noted for their beautiful coloration, wing pattern and flight. They are popularly known as raja, nawab, baron, duke, duchess, commodore, sergeant, sailor, map, oakleaf, pansy, leopard, lacewing etc.

Nymphalid butterflies vary considerably in color being bright to brilliant. In size they range from 25-130 mm. Sexual dimorphism is present in some species while in others seasonal forms are present. A few species are known as mimics of danaiids and there by gain protection. There are over 400 species of nymphalids in the Indian sub region, of which 48 species occur in Kerala. The common bush brown (*Modusa procris*), the common sailor (*Neptis hylas*), the great egg fly (*Hypolimnas bolina*), the yellow pansy (*Junonia hierta*), the lemon pansy (*J. lemonias*), the baron (*Euthalia garuda*) and the tamil yeoman (*Cirrochroa thais*) are some of the species commonly found in Kerala.

The immature stages feed on *Tamarindus indica*, *Caesalpinia* sp., *Adenantha pavonina*, *Albizia* spp., *Mangifera indica*, *Anacardium occidentale*, *Haldina cordifolia*, *Xylia xylocarpa*, *Olea dioica* etc.

## **Lycaenidae**

They are predominantly blue in colour and are generally small in size. The immature stages mostly feed on Acanthaceae (*Strobilanthus* sp.), Amaranthaceae (*Amaranthus* sp.), Cycadaceae (*Cycas* sp.), Leguminosae (*Abrus* sp., *Albizia* sp., *Cassia* sp., *Xylia* sp.), Utricaceae (*Ficus* sp.) and Verbanaceae (*Clerodendron* sp. and *Lantana* sp.). There are about 650 species in India, of which 35 are known from Kerala which include the gram blue (*Euchrysops cnejus*), the common cerulean (*Jamides celeno*), the metallic cerulean (*J. alecto*) and the common pierrot (*Castalius rosimon*).

## BUTTERFLY FARMING

Butterfly farming is closely linked with establishment of 'butterflyhouses'. Perhaps the roots of this technique are in Britain where the 'butterflyhouse' exhibit has grown from two different lepidopteral industries: first from breeders of European butterflies who began breeding exotic tropical species and secondly from the silk industry where some breeders began displaying certain exotic silk moths to visitors.

From the merger of these two activities and the provision of large glass enclosures, there developed the 'butterflyhouse' - which emerged as a public attraction in the 1960s. The first butterfly farm- the Guernsey Butterfly Farm was established in the U.K. in the 1960s. The 1980's saw a massive expansion of the business with over 40 facilities being reported by 1986 . A survey conducted in 1987 in 26 butterfly houses in UK has indicated that 306 species of butterflies were used in the butterfly houses with 26-75 species being kept at any one time (Collins, 1987).

The concept of captive breeding is also relevant in species conservation programmes. On occasions when a species is threatened with extinction due to destruction of its habitat and absence of other suitable sites to which it could be transferred , there is a need to maintain stocks in captivity for use in future reintroductions (Morton, 1983). At Wood Walton Fen in Britain, for example, a free living population of the Dutch race of the large copper butterfly *Lycaena dispar batavus* is supplemented annually by specimens reared in captivity (Nature Conservancy Council, 1977). The concern for the conservation of butterflies has led to the formation of the IUCN specialist group on butterflies.

Thus, butterfly farming is a novel technique which simultaneously meets commercial demands for specimens with out affecting wild populations, puts cash into the local economy and (probably most important) fosters local interest and commitment to the conservation ethic. As a commercial enterprise and as a conservation oriented programme, butterfly farming is highly promising. Butterfly farming is beneficial in conservation of tropical forests and also provides rural economics with income (Pearson. 1992).

## MATERIALS AND METHODS

An existing insectory (8m x 5m x 5m) covered with fine steel netting and provided with a double door was made use of for rearing purpose (Figs. 1a and 1b). Within the cage appropriate larval host plants were maintained to facilitate oviposition by the butterflies released. In addition to this, provision was also made to keep diluted sugar solution for sustaining butterflies.

An area of approximately 0.5 ha surrounding the insectory was landscaped so as to produce different habitats such as dense vegetation (Figs. 2a & 2b), open areas, bushes etc., in order to attract local butterflies having different habitat preferences. The species planted in this area include *Aristolochia indica*, *Murraya koenigi*, *Crotalaria retusa*, *Lantana camara*, *Clerodendron capitatum*, *Cassia* sp., as well as *Citrus* sp., which serve as host plants of several butterflies like *Pachliopta aristolochiae*, *Papilio demoleus*, *Danaus genuta* and *Tirumala limniace leopardus* (Appendix-I). Regular observations were also made on butterflies visiting various plants.

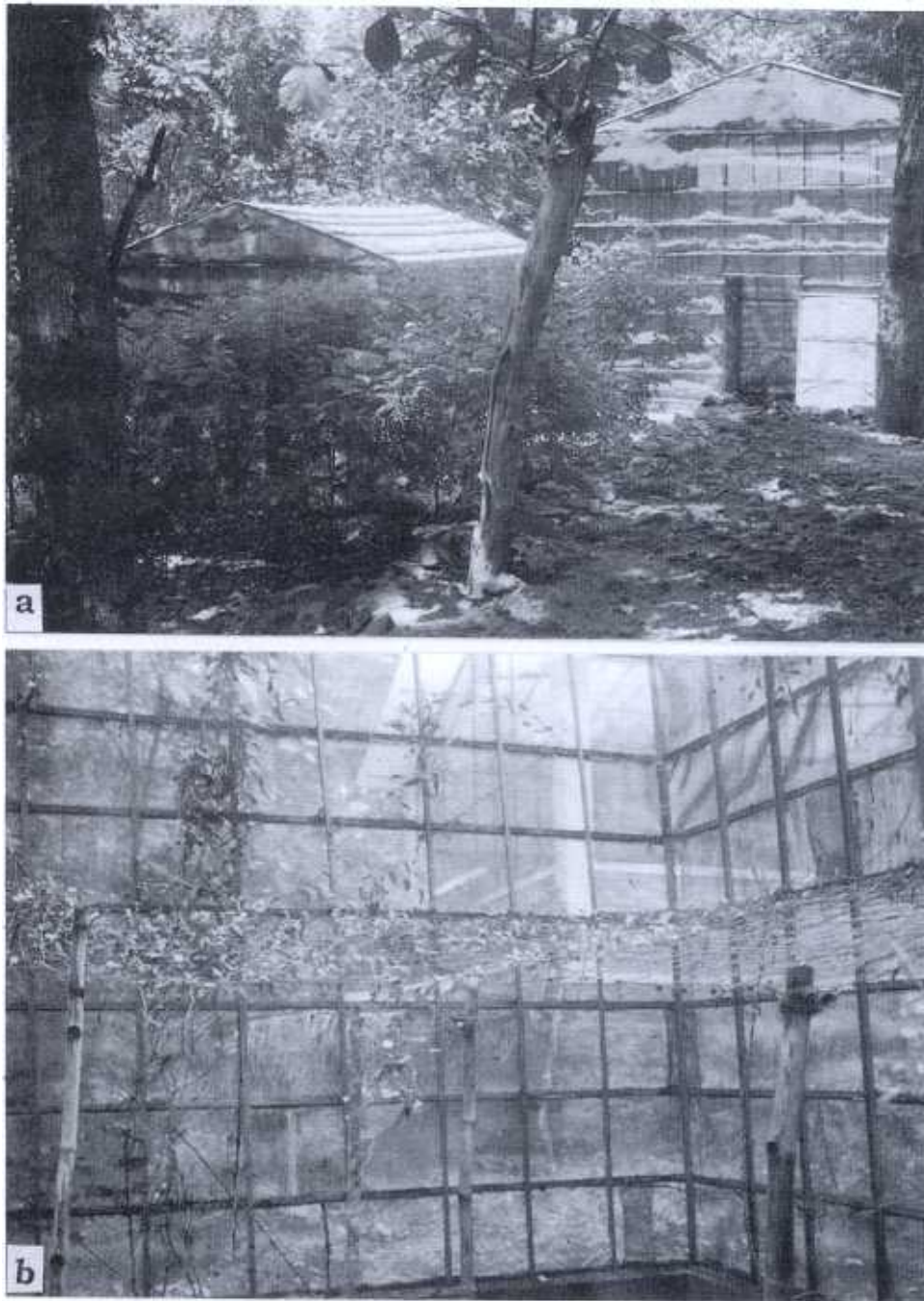


Fig. 1. a. View of the butterfly house in KFRI campus  
b. Interior of the butterfly house with *Aristolochia indica* grown for breeding the common rose, *Pachliopta aristolochiae*

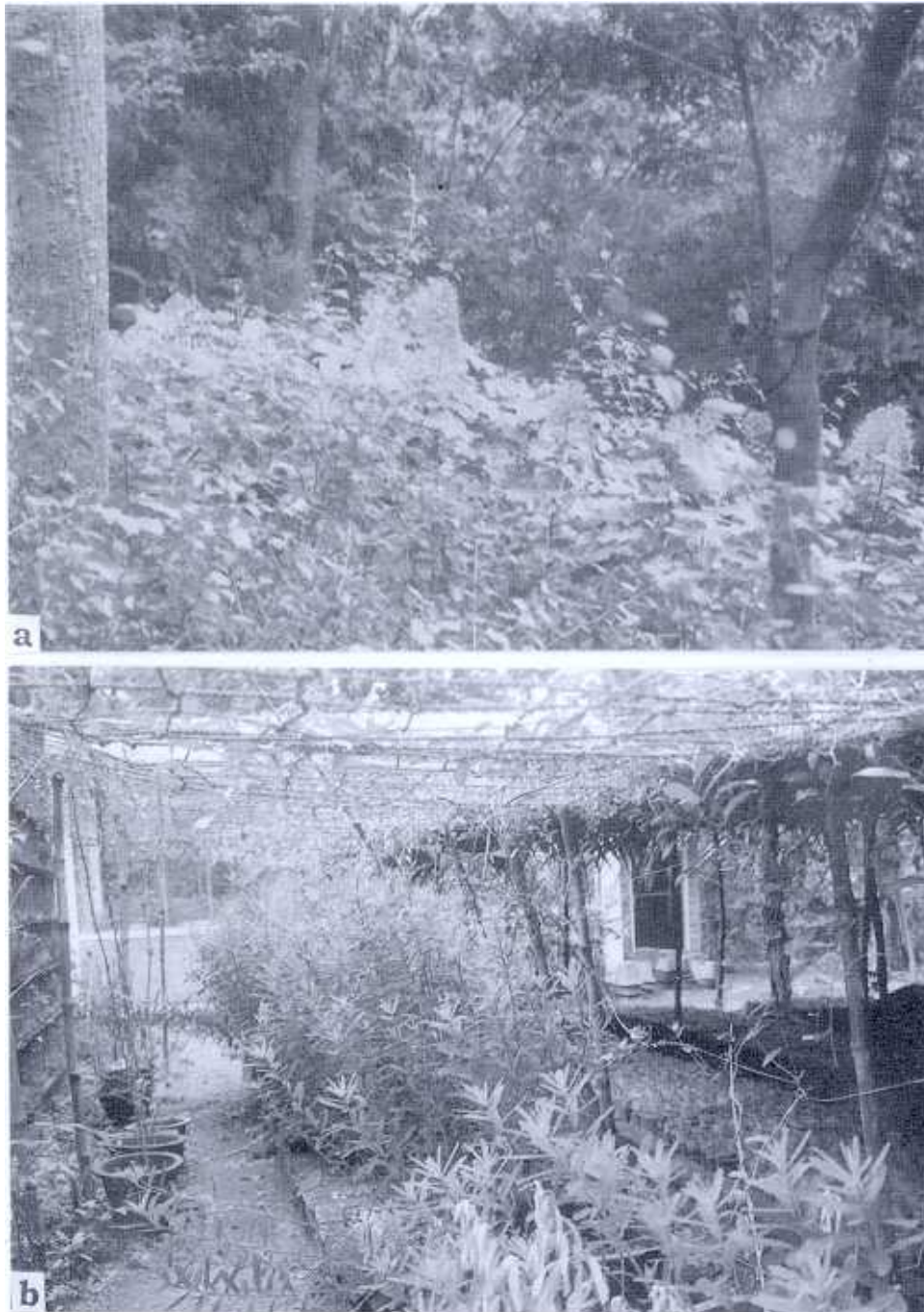


Fig. 2. a. View of the butterfly garden in KFRI campus containing flowering plants like *Lantana camara* and *Clerodendron paniculatum*  
b. *Crotalaria* grown into hedges for attracting danaine butterflies

## RESULTS AND DISCUSSION

It took nearly two years for establishing the plants in the insectory and in the surrounding area. With the establishment of plants, various butterflies started to frequently visit the area leading to the formation of a small local population.

### BUTTERFLIES OBSERVED IN THE BUTTERFLY GARDEN

A list of butterflies, with details of their number and occurrence pattern are given in Table 1. Altogether, 11 species of butterflies belonging to the families

**Table 1. List of butterflies recorded from the butterfly garden established at Peechi**

Family/species	Plant host	Remarks
<b>Papilionidae</b>		
<i>Papilio paris</i> Moore	<i>Clerodendron paniculatum</i> , <i>Lantana camara</i>	1 or 2 individuals visiting flowerheads in the morning and afternoon
<i>P. polymnestor parinda</i> Moore	"	"
<i>P. polytes</i> Cramer	"	"
<i>P. demoleus</i> Lin.	"	"
<i>Troides minos</i> Cramer		"
<i>Tros hector</i> Lin.		Several individuals feeding at flowers
<i>Pachliopta aristolochiae</i> Fb.		
<b>Danaidae</b>		
<i>Danaus genyita genyita</i> Cramer	<i>Crotalaria retusa</i>	20 to 30 butterflies aggregating on foliage
<i>Tirumala limniace leopardus</i> Butler		
<i>Euploea core core</i> Cramer		
<b>Pieridae</b>		
<i>Eurerna blanda</i> Boisd.		A few butterflies (2-3 nos.) frequenting flower heads of <i>L. camara</i> and <i>Cassia</i> sp.

Papilionidae, Danaidae and Pieridae were observed in the study area. The flower heads of *Clerodendron paniculatum* and *Lantana camara* were found to be very much preferred by butterflies belonging to Papilionidae and Pieridae. The danaids *Danaus genivitta*, *Tirumala limniace* and *Euploea core* were found to aggregate in large numbers on the foliage of *Crotalaria retusa*. These butterflies were observed to lacerate the leaf surface by scratching with the front tarsal claws and then lick the exuding sap. The lacerated leaves later withered. Up to 20 butterflies were noted to feed gregariously on a single plant and heavy incidence of butterflies led to the wilting of plants. Danaid butterflies are already reported to feed on plants containing alkaloids. Caterpillars of an arctiid *Argyria* sp. were found to develop on foliage of *C. retusa* occasionally causing die-back of crotalaria seedlings (Figs. 3a & 3b).

An evaluation of the establishment of various butterflies in the study area was also made by examining the larval host plants for the presence of their immature stages. The hedges of *Aristolochia indica* contained eggs, larvae and pupae of the common rose *Pachliopta aristolochiae* (Fig. 4a) and of the southern bird-wing *Troides minos* (Fig. 4b). Similarly, the immature stages of the lime butterfly *Papilio demoleus* (Fig. 5a) was noted on *Citrus* sp., and *Murraya koenigi*; *Moduza procris* on *Mussaenda frondosa* and *Euploea core* on *Ficus glomerata*. Observations were also made on the occurrence of butterflies in the garden during the various months (Table 2). Data gathered has shown that, in general, the number of butterflies present was high during the months September to December. The butterflies showed a reduction in population during in summer (March-May).

Seven species of papilionids including *Papilio paris*, *P. polymnestor*; (Fig. 5b) *P. demoleus*; the danaids *Pachliopta aristolochiae*, *Parantica nilgiriensis* and *Euploeacore* were observed quite frequently in the garden. The pierids *Eurema hecabe* and *Catopsilia pomona* (Fig. 6a and 6b) were also found in small numbers feeding at the flowers of *Lantana camara* and *Cassia* sp.

## **BUTTERFLIES RELEASED IN THE FIELD CAGE**

Preliminary attempts to farm butterflies in captivity were made by releasing both the adult and immature stages of certain commonly available butterflies like *Troides minos*, *Papilio demoleus* and *Pachliopta aristolochiae* in the insectory, where appropriate larval host plants of each of these butterflies (*Citrus* sp., *Aristolochia indica* etc.) have been established. Water as well as diluted sugar solution were kept in separate containers for feeding the butterflies.



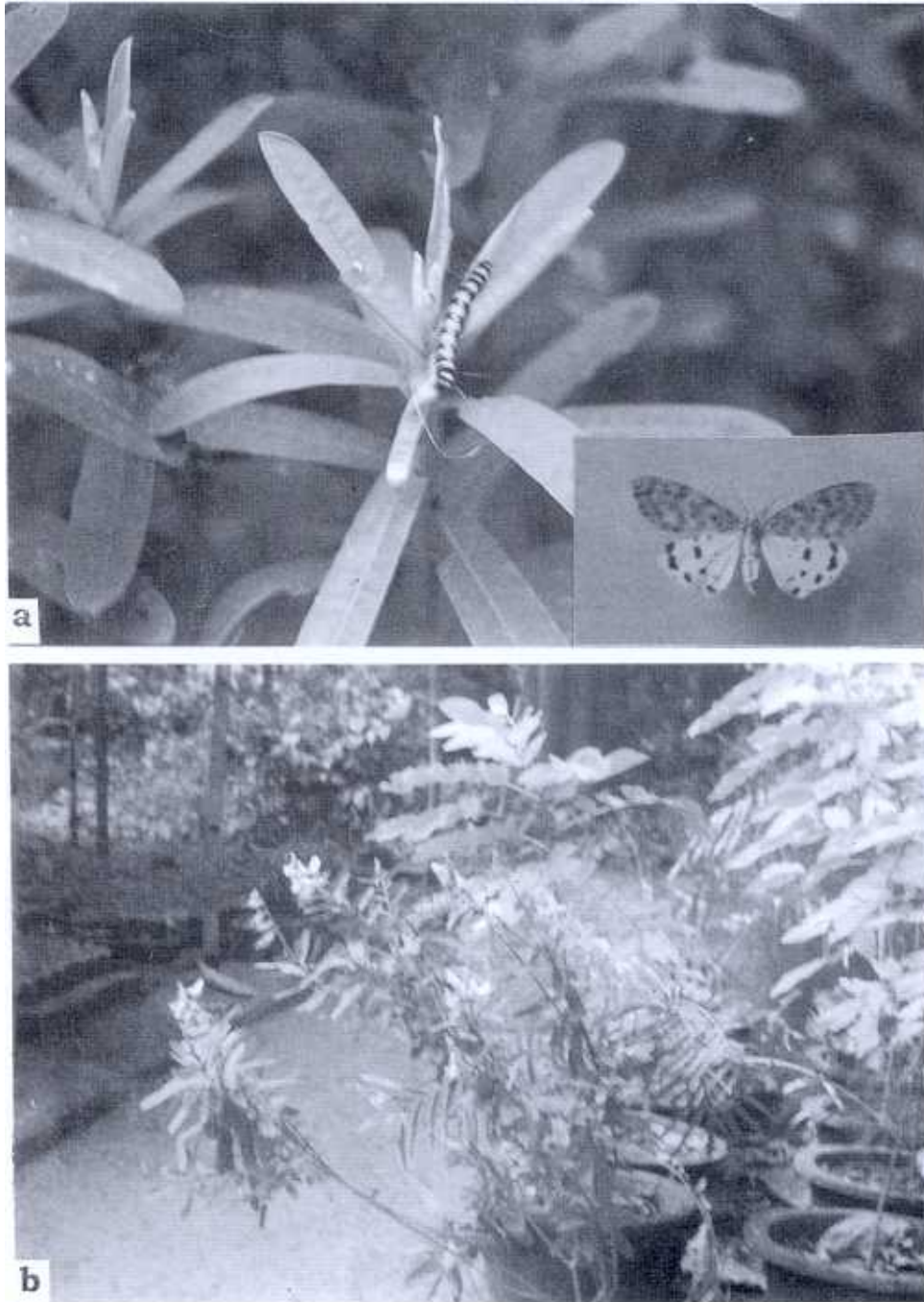


Fig. 3. a. Close-up of *Crotalaria retusa* showing a caterpillar of *Argyria* sp. (Arctiidae). Inset: The moth  
b. *Crotalaria retusa* in bloom. Both leaves and flowers attract and sustain various danaine butterflies

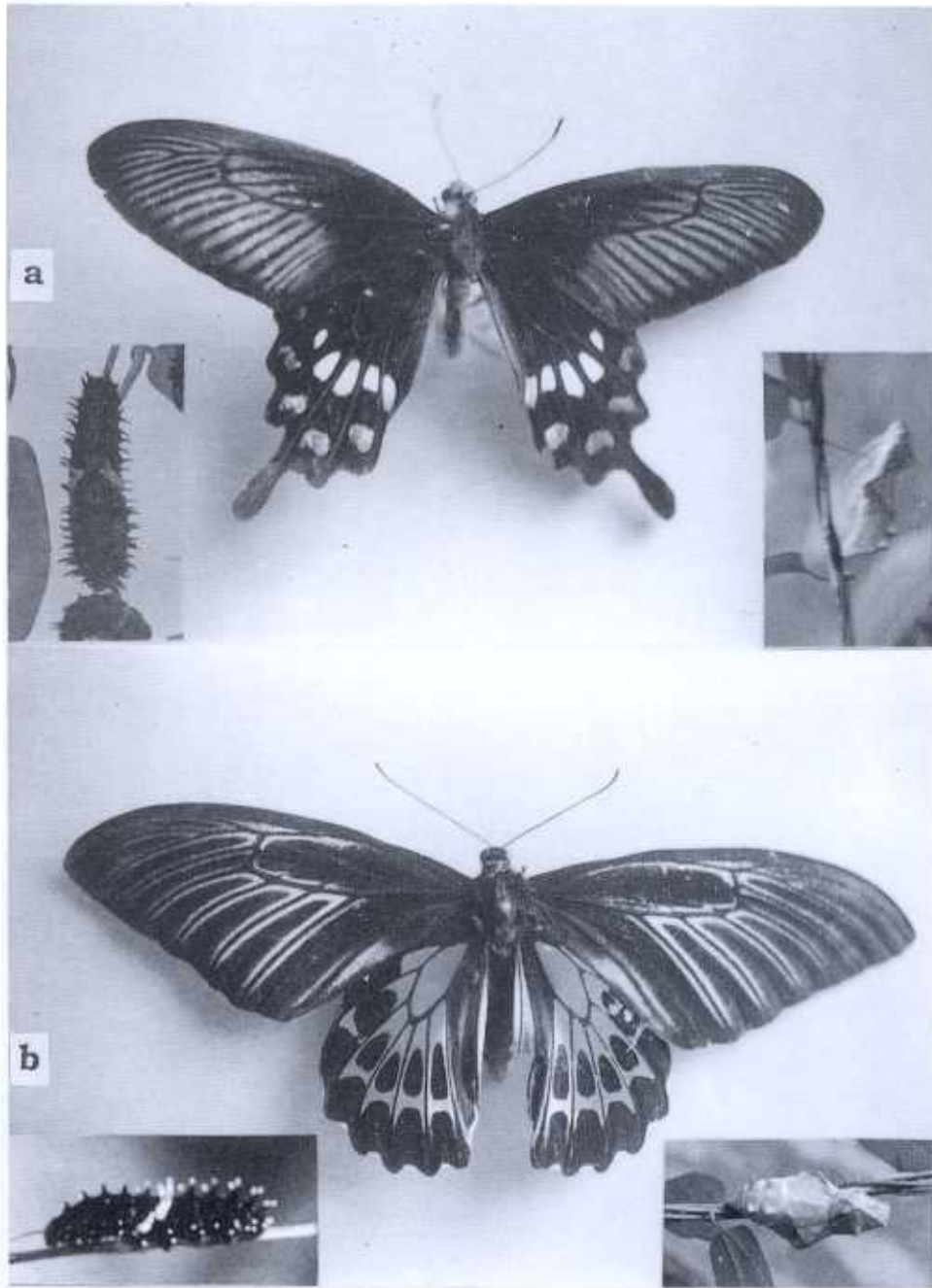


Fig. 4. a. The crimson rose, *Pachliopta aristolochiae*. Inset: Larva and pupa  
b. The southern bird wing, *Troides minos*. Inset: Larva and pupa

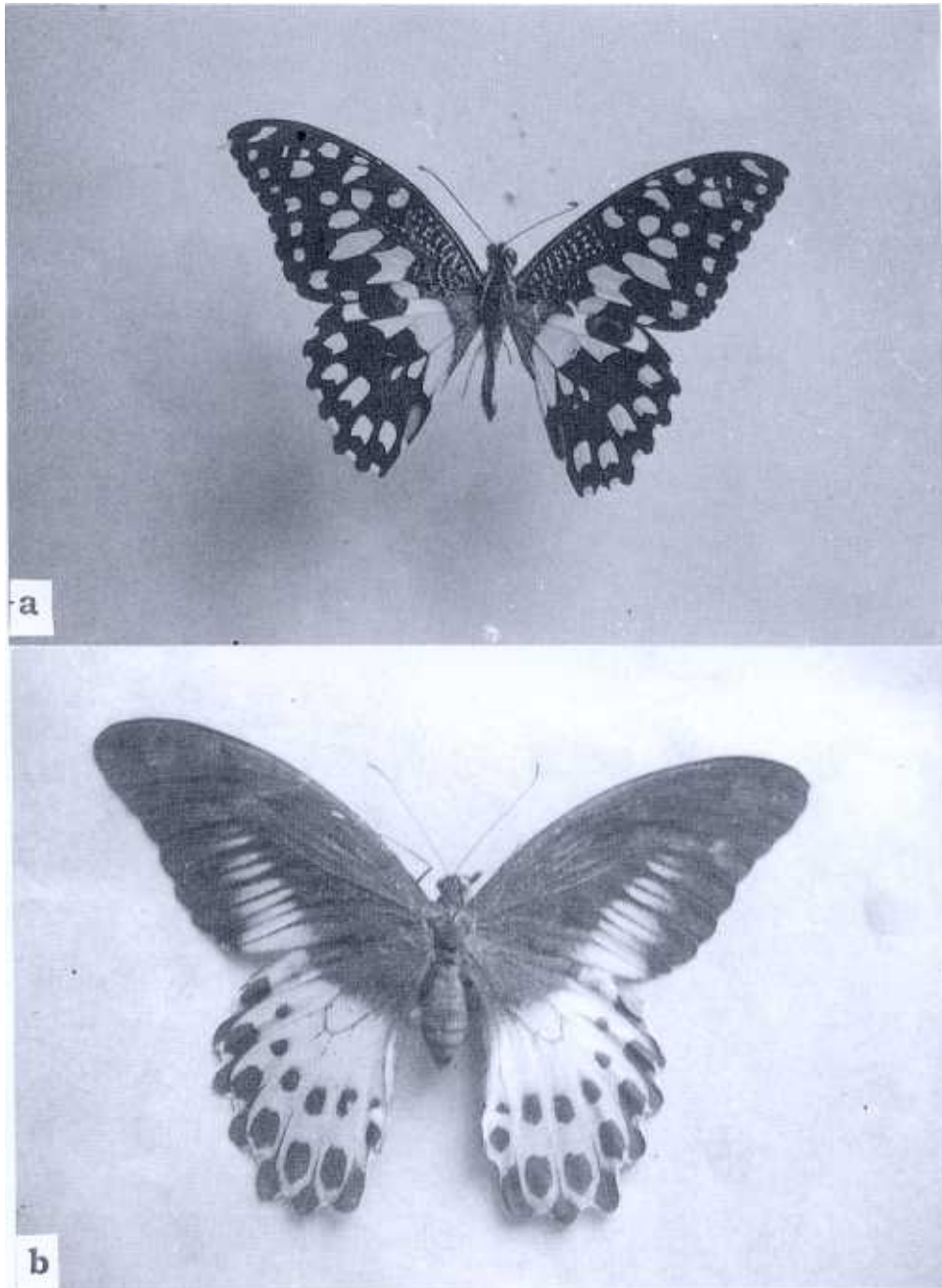


Fig. 5. a. The lemon butterfly, *Papilio demoleus*  
b. The blue mormon *Papilio polymnestor*

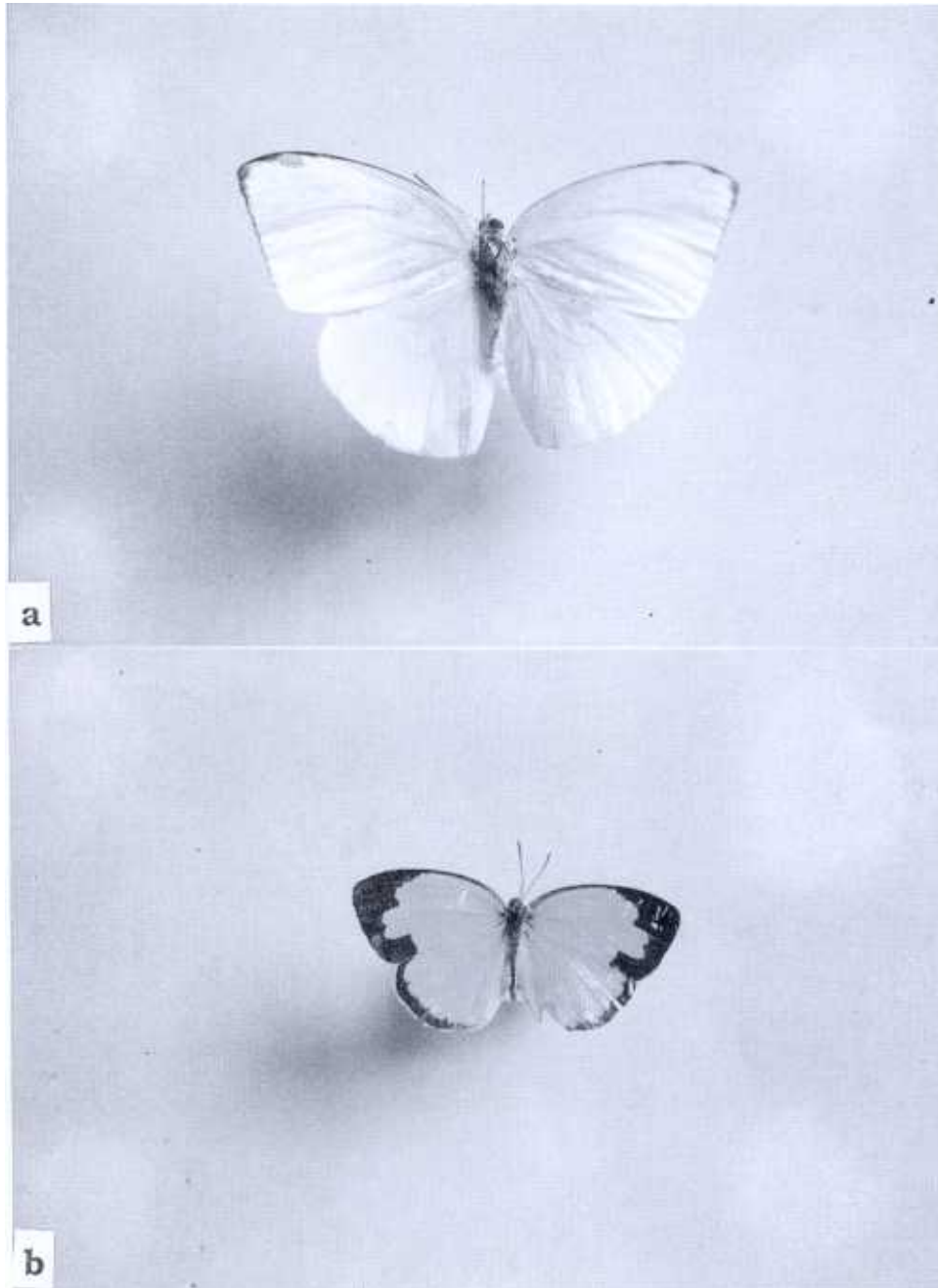


Fig. 6. a. The white, *Catopsilia pomona*  
b. The grass yellow, *Eurema hecabe*

**Table 2. Butterflies observed in the garden during various months**

year and month of observation	Butterfly species	Stage	Host plant
1996			
May			
June	<i>Papilio demoleus</i>	Larava & adult	Citrus sp.
July	<i>Troides minos</i>	Larva	<i>Aristolochia indica</i>
	<i>Euploea core</i>	Adult (aggregation)	<i>Crotalaria retusa</i>
	<i>Tros hector</i>	Larva	<i>A. indica</i>
August	<i>Troides minos</i>	Adult	<i>Clerodendron capitatum</i>
September	<i>Euploea core</i>	Adult (aggregation)	<i>Crotalaria retusa</i>
	<i>Parantica nilgiriensis</i>	Adult (aggregation)	<i>Crotalaria retusa</i>
October	<i>Euploea core</i>	Adult (aggregation)	<i>Crotalaria retusa</i>
	<i>Parantica nilgiriensis</i>	Adult (aggregation)	<i>Crotalaria retusa</i>
November	<i>Pachliopta aristolochiae</i>	Adult (aggregation)	<i>Aristolochia indica</i>
	<i>Papilio demoleus</i>	Larva & adult	<i>Citrus sp.</i>
	<i>Papilio paris</i>	Adult	<i>Clerodendron paniculatum</i>
December	<i>Pachliopta aristolochiae</i>	Adult (aggregation)	<i>Aristolochia indica</i>
1997			
January	<i>Pachliopta aristolochiae</i>	Larva	<i>Aristolochia indica</i>
February	<i>Pachliopta aristolochiae</i>	Larva	
	<i>Modusa procris</i>	Larva	<i>Mussaenda frondosa</i>
March	<i>Pachliopta aristolochiae</i>	Larva & adult	<i>Aristolochia indica</i>
April	"	"	"
May	"	"	"
June	<i>Parantica nilgiriensis</i>	Adult (aggregation)	<i>Crotalaria retusa</i>
	<i>Euploea core</i>	"	"
July	<i>Parantica nilgiriensis</i>	"	"
	<i>Euploea core</i>	"	"

The butterflies thus released survived for about a week but no instance of mating and egg laying was noticed. Perhaps the area available within the insectary was not sufficient to induce mating response. The butterfly farms established in Malaysia had large areas (covering over 100m<sup>2</sup>) under enclosure so as to provide as much natural space as possible. Even in this case,

the natural build up of butterflies on the host plants was not satisfactory and butterflies had to be introduced into the cage at intervals in order to maintain the population within the cage. Details of butterflies released into the cage area given in Table 3.

**Table 3. Details of butterflies released in the insectory**

Species	Stage	No. and date of release	Remarks
<i>Tros hector</i>	Larva	7 Nos. 19-7-96	Pupation from 21-7-96 to 23-7-96. Adult emergence from 1-8-96 to 3-8-96
	Adult	4 Nos. 22-7-96	Found dead on 26-7-96
<i>Euploea core</i>	Larva	1 No. 30-10-96	Pupated on 5-11-96. Adult emergence on 15-11-96
<i>Pachliopta aristolochiae</i>	Larva	10 Nos. 24-10-96	Pupation from 28- 10-96 to 30- 10-96 Adult emergence from 5-11-96 to 8-11-96
<i>Troides minos</i>	Larva	2 Nos. 2-7-97	Pupated on 15-7-97. Adult emergence on 21-7-97
	Pupa	1No. 21-7-97	Adult emergence on 28-7-97

## CONSTRAINTS FOR BREEDING BUTTERFLIES IN CAPTIVITY

Being a pilot study, attempts were made to evaluate the various requirements for the successful breeding of butterflies in field cages. The constraints faced in this study are discussed below.

### Out door cage

In the present study, an insectory in the KFRI campus was made use of. Although butterflies were reared in the laboratory in rearing bottles and occasionally in small cages, the dimension of cage was found to be important for the success of the breeding programme particularly when the intention is to maintain butterflies in a natural

### Standardised strategy for breeding

General information on the biology and ecology of butterflies to be bred, natural mortality factors as well as availability of appropriate larval and adult

host plants are essential for the success of any breeding programme. In the present study, selected butterflies were initially bred in small containers in the laboratory prior to release in the field cage. Predation by lizards and infestation by micro-organisms were the major reasons for the decline of population maintained in the field cage. Instances of 'inbreeding' leading to a loss of vigor due to loss of heterozygosity and fixation of homozygous alleles have been reported in continued breeding programmes. Since most species of butterflies have more than one host plants, care should be taken to select the most appropriate host.

## **GENERALISED PLAN OF A BUTTERFLY HOUSE**

A typical butterfly house should have facilities to breed various species in captivity and also to display preserved material in order to create public awareness on species conservation (Fig. 7). A detailed account of facilities required for a butterfly house are given below.

### **Display area**

Insects particularly butterflies form excellent materials for exhibits and hence should be displayed in the front area. The exhibits should be so organised as to give general information on species diversity and its role in ecosystem functioning. Both native and exotic fauna may be exhibited (Fig. 8). Commercial products made out of insects, souvenirs etc., also may be exhibited in this area.

### **Sales counter**

Souvenirs and curios made out of insects, plastic models, charts, books, stickers, posters, paintings, insect study and collection kits, T-shirts, badges, bags etc.. form excellent material for sale. The sale proceeds from such items together with gate collections will bring good revenue to make the butterfly house self supporting.

### **Butterfly release area**

The butterflies are generally reared in separate rearing rooms either on their larval host plants or on artificial diets. The butterflies are subsequently left in the release area which is open to the public.

The release area should be developed with care so as to offer conditions suited for the butterflies released (Fig. 9). It may be either a simple net house or a

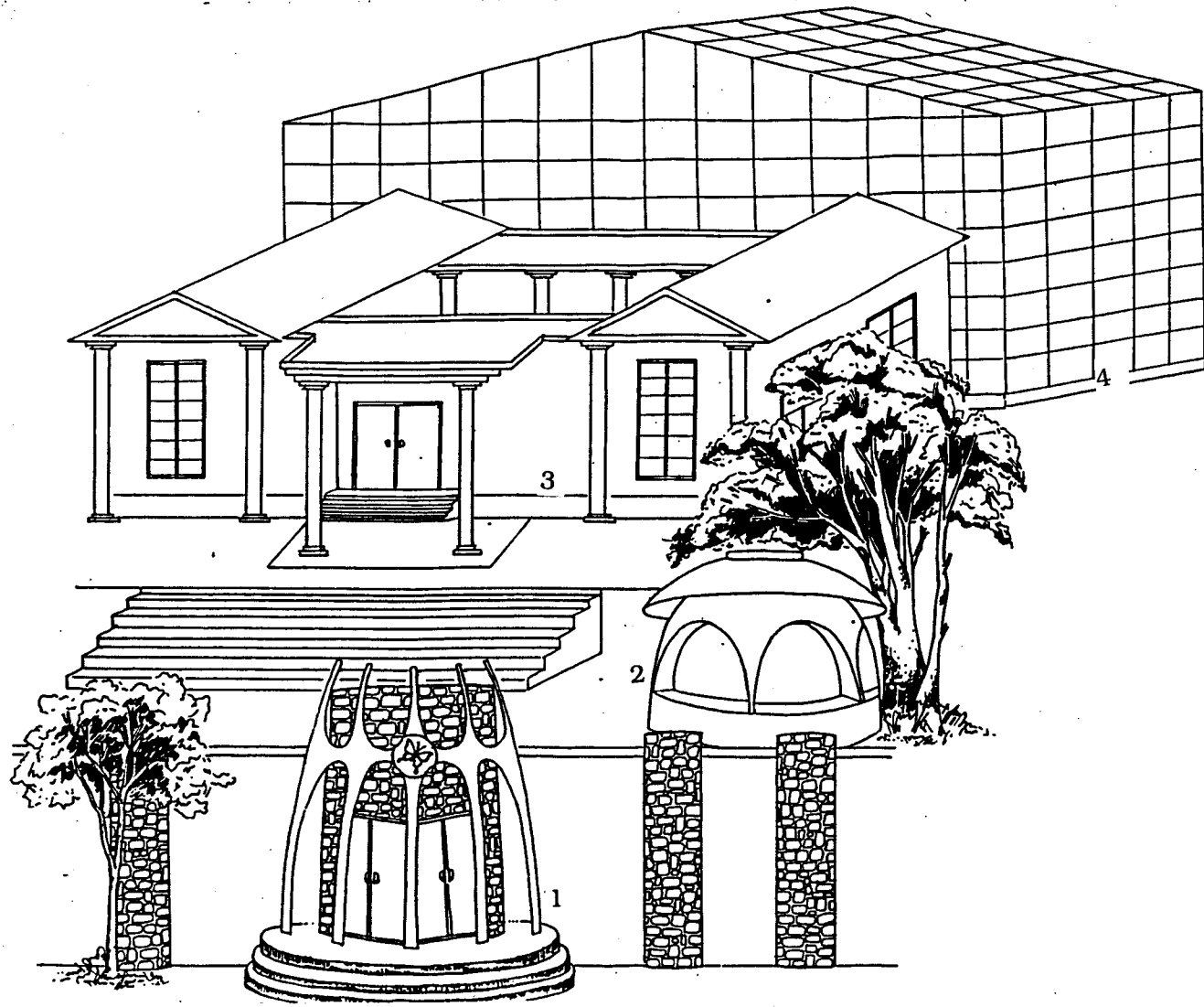


Fig. 7. General plan of a typical butterfly house.  
1. Gate, 2. Caffeteria, 3. Sales counter-cum-exhibit  
area, 4. Butterfly release area



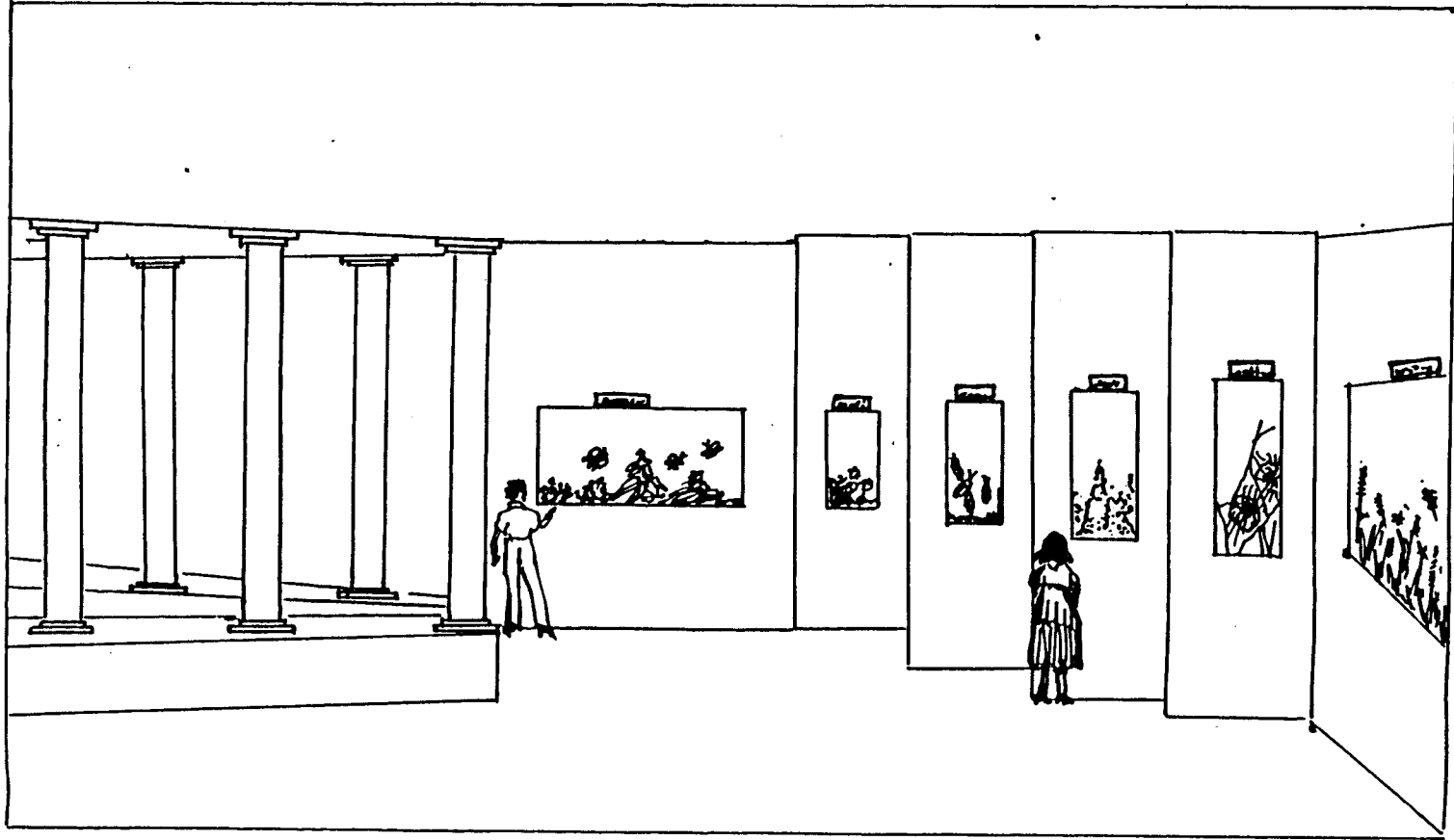


Fig. 8. Display area in a typical butterfly house

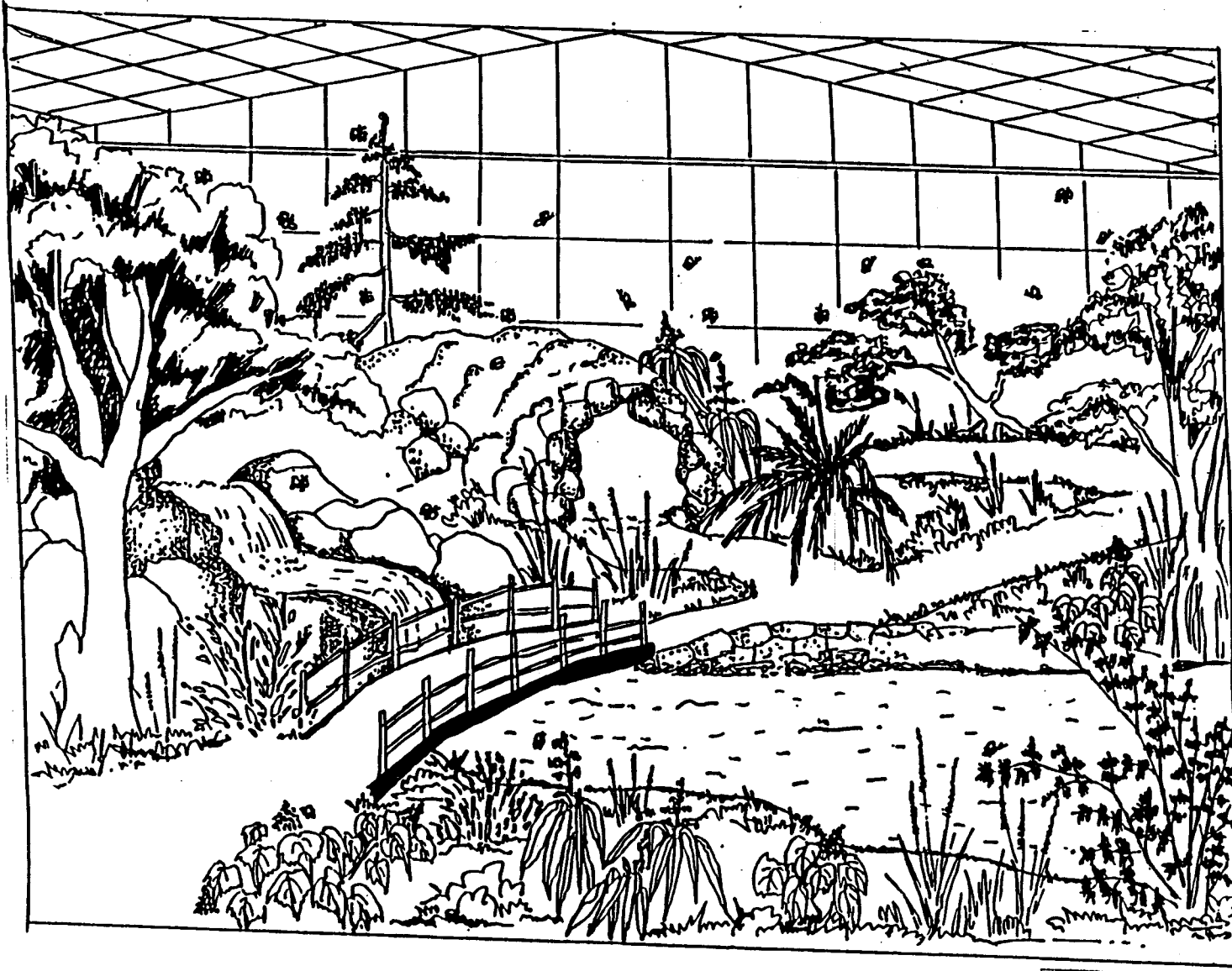


Fig. 9. Inside a butterfly house. Note the properly land-scaped interior offering diverse habitats

glass house with adaptations to regulate temperature, humidity, air movement and day length. Temperature and relative humidity are the most important factors to be taken care of. Temperature ranging from 20-30° seems to be preferable for most tropical butterflies. Excessive humidity may lead to development of fungal attack. Most species also thrive best at a relative humidity exceeding 90 percent. In addition to these, slight air currents will stimulate them to move about leading to a high frequency of feeding and courtship.

In order to offer conditions for mating and egg laying, appropriate larval and adult host plants should be kept in the release area. As nectar and pollen from this source alone may not be sufficient, provision may also be made for providing artificial food. Generally, sugar solution as soaked cotton wads is kept. In some butterfly houses, the solution is kept in a tube. Colored 'corolla' or plastic flowers are kept around the tube to attract butterflies. Powdered pollen available from bee keepers may also be kept on the 'corolla'. Besides these, over-ripe fruits dusted with sugar, carrion, dung etc., may also be used depending on the species. Sufficient care should be taken to keep the solution, containers and 'corolla' disease free. For this, these may be rinsed in 5% sodium hypochlorite solution before use.

Most of the commercial butterfly houses in the west are walk through heated glass houses fitted with special halide lamps for lighting as daylight is completely cut off. For tropical conditions such arrangements may not be required. Butterfly houses in Malaysia for example, are only enclosures with simple provisions for air regulation and humidity control and do not use much sophisticated techniques as in the west.

## **PREPARATION OF CURIOS**

In many countries like Papua New Guinea, preparation of 'curios' using dead insects has grown into a cottage industry. The wings of many butterflies are very attractive and hence even bits or fragments of wings can be made use of in the preparation of various art works. There is strong criticism in the preparation of such articles mainly due to the fear that this may lead to exploitation of local fauna and may affect even the survival of the species. What is suggested here, is to make use of dead insects that become available in an insectary during routine breeding programmes. It is strongly recommended that no attempt should be made to rear the insects merely for commercial utility. Some general techniques for processing dead butterflies or fragments are discussed below.

## **Dry mounts**

Insects that are not mutilated can be spread on boards and allowed to dry in the air or in the oven at 50-55°C. Such dried material will retain its shape and can be assembled on a board and framed or retained in attractive display boxes. Appropriate fumigants -like naphthalene pellets - may be kept inside the boxes to prevent attack by small insects like psocids. Another problem with dry mounts is fungal attack during wet seasons. If the box is air tight, this problem can be managed to some extent.

## ***Plastination***

This is a technique used for preserving biological specimens by embedding in a polyester or epoxy resin. For getting best results the material used should be fresh. The procedure for plastination is as follows. At first, the material is subjected to dehydration by freeze drying in vacuum. This can be achieved by leaving the material in a desiccator to which is attached a vacuum pump. Treatment in cold acetone at -25°C will also enhance dehydration. The material is then soaked in a polyester resin to replace the tissue fluid. As the chemical will get polymerised at room temperature, care should be taken to keep the material in the desired shape and after polymerisation the shape can not be altered. The impregnated material is then treated with a hardener or catalyst to hasten the drying of the resin.

## ***Mica-ge embedding***

This technique is similar to plastination, the only difference being that the material is embedded in a thick film of polyester resin. For this, a glass or mica container with detachable sides is taken into which the resin is slowly added to about 1/4 th of its depth. The dried and set material to be embedded is then kept over the surface. The resin is continuously added till the whole material is thoroughly embedded. While adding the resin care should be taken to prevent formation of air bubbles. Also, the resin should be added with least disturbance so that the material will not lose its natural shape. In order to hasten quick setting of the resin, a catalyst is also added after embedding. When completely dry, the block may be polished using machine.

## CONCLUSIONS

Captive breeding of butterflies has economic as well as ecological implications as detailed below.

### AS A BUSINESS

In many countries such as Papua New Guinea, Britain and Malaysia, butterfly farming has proven to be a sound, economically viable rural industry. Papua New Guinea's is a successful model that can be readily adopted to other tropical developing countries (Anon., 1983; Hutton, 1985). In that country, there are several village farming and collecting units particularly in areas that are handicapped for the usual agricultural activities, or livestock maintenance. Butterflies are bred in these areas with minimal forest disturbance.

Current estimates indicate that there are between 50 and 60 butterfly houses attracting five million visitors annually, in Britain. Millions of butterflies are sold as specimens or as displays. It has been stated that the gate collections alone exceeded £ 5 million in Britain (Collins, 1987).

### AS A TOOL IN CONSERVATION AND ENVIRONMENTAL EDUCATION PROGRAMMES

Invertebrates which form the majority of the life forms on earth have been ignored until recently in zoo education and conservation programmes. During the past decade 'butterfly houses' have become very popular in several countries attracting millions of visitors.

In most environments, invertebrates are more accessible than vertebrates and they offer insights into the dynamics of ecological processes that can raise public awareness and enjoyment (Collins, 1987). The butterflies which are very colorful and attractive rival the most beautiful of the tropical fish, flowers and birds. Butterfly houses can thus promote awareness of the vital ecological roles played by invertebrates and particularly the need to conserve invertebrates as important natural resources.

Captive breeding and maintenance of stock for reintroduction is a method applied in vertebrate conservation. The possibility of invertebrate

tion on the same lines have been suggested by several workers but largely ignored (Cooper, 1986, Morton, 1983). Pyle (1976) states that such measures should be regarded as the last resort. Introduction of species into new areas had also aroused much controversy. Such action, it is stated, would be justified only when a species has become locally extinct or when a species is facing threat of extinction due to habitat destruction.

Several instances of reintroduction of butterflies are known such as the introduction the large copper butterfly *Lycaena dispar* Haw. *batavus* Obth. into Woodwalton Fen during 1927 to replace *L. dispar* Haw. which became extinct towards the middle of the 19th century (Nature Conservancy Council, 1977). Similarly, the heath fritillary *Mellicta athalia*, which became extinct in Essex in about 1890 was reintroduced in the Hadleigh Woods with success (Luckens, 1980). Similarly attempts were also made to introduce the swallow-tail butterfly *Papilio machaon britannicus* at Wicken Fen in 1975. However, such introductions are terribly expensive since all infrastructural facilities for breeding and maintenance of populations have to be built up. Establishment of a Captive Breeding Center is therefore very essential for the success of such programmes.

Perhaps the most important step in conservation should be legislation against collecting for trade particularly with regard to species that are rare. For this, information on the local fauna, the population density of various species as well as the various natural mortality factors affecting species survival should be known. Only detailed faunistic surveys can bring out these details.

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## Appendix-1

**Butterflies recorded from southern India with notes on their distribution and host range  
(Wynter-Blyth, 1957; D'Abrera, 1982-1986; Larsen, 1987, 1988)**

Order\Species	Status	Distribution	Host plant
<b>DANAIDAE Idea (Tree Nymphs)</b>			
<i>Idea malabarica</i> Moore	Common	S. India	<i>Aganosma cymosa</i>
<b>Danais (Tigers)</b>			
<i>Parantica aglea</i> (Cramer)	Fairly common	Sri Lanka, S. India, to Poona	<i>Tylophora</i> sp., <i>Cryptolepis buchananii</i> , <i>Calotropis gigantea</i>
<i>P. nilgiriensis</i> Moore	Common	Nilgiris and hills to the south	
<i>Danaus limniace</i> Cramer	Very common	India, Sri Lanka	<i>Wattakaka volubilis</i> , <i>Calotropis</i> , sp., <i>Marsdenia tenacissima</i>
<i>Tirumala septentrionis dravidarum</i> Fruh.	Very common	S. India, Sri Lanka	Asclepiads; <i>Vallisneria spiralis</i>
<i>D. genutia genutia</i> Cram.	Fairly common	India, Sri Lanka	<i>Cynanchum dalhousiae</i> , <i>Ceropegia intermedia</i> , <i>Rhaphistemma pulchellum</i>
<i>D. chrysippus</i> (Lin.)	Very common	Sri Lanka, India	<i>Calotropis gigantea</i> , <i>C. procera</i> , <i>Asclepias curassavica</i>
<b>Euploea (Crows)</b>			
<i>Euploea core</i> (Cramer)	Very common	Sri Lanka, India	<i>Holarrhena pubescens</i> , <i>Nerium oleander</i> , <i>Ichnocarpus frutescens</i> , <i>Streblus asper</i> , <i>Ficus bengalensis</i> , <i>F. racemosa</i> , <i>F. indica</i> , <i>F. religiosa</i> , <i>Hemidesmus indicus</i> , <i>Cryptolepis elegans</i>
<i>E. sylvester coreta</i> (Godart)	Locally very common	S. India, Sri Lanka	<i>Ichnocarpus frutescens</i>
<i>E. klugii kollari</i> Felder and Felder	Rare	Sri Lanka, S. India, Bengal and Konkan	<i>Ficus hispida</i>
<b>SATYRIDAE</b>			
<b>Mycalesis (Bushbrowns)</b>			
<i>Mycalesis anaxias</i> Hewitson	Locally common	Hills of S. India, Coorg	Grasses
<i>M. perseus</i> (Fb.)	Very common	Sri Lanka, India	S. Grasses
<i>M. mineus</i> (Lin.)	Very common	Sri Lanka, Peninsular India	Grasses
<i>M. visala</i> Moore	Locally abundant	S. India	Grasses
<i>M. subdita</i> (Moore)	Locally common	Sri Lanka, Nilgiris, Madras to Orissa, Bangalore	Grasses
<i>M. khasia</i> Evans	Common	Palnis, Shevaroys, Nilgiris, Wynad, Coorg	Grasses
<i>M. adolphet</i> (Guerin)	Fairly common	Nilgiris, Coorg	Grasses
<i>M. oculus</i> Marshall	Common	Hills of S. India (South of Nilgiris)	Grasses
<i>M. patnia</i> Moore	Common	Sri Lanka, S. India to Mysore and North Kanara	Grasses
<b>Lethe (Tree browns)</b>			

<i>Lethe europa</i>	Rather rare	S. India	
<i>L. rohria</i>	Common	Sri Lanka, S. India	Bamboo
<i>L. drypētis</i>	Common	Sri Lanka, S. India	
<b>Ypthima</b> (The Rings)			
<i>Ypthima chenui</i>	Common	Coorg, Hills of S. India	
<i>Y. ceylonica</i>	Locally abundant	Sri Lanka	
<i>Y. avanta</i>	Common	Sri Lanka, Peninsular India	Grasses
<i>Y. philomela</i>	Locally common	Nilgiris, Wynad	
<i>Y. baldus</i>	Very common	S. India	
<b>Zipoetis</b> (The Catseyes)			
<i>Zipoetis saltis</i> Hewitson	Local	Slopes of W. Ghats	<i>Lantana camara</i>
<b>Orsotriaena</b> (The Nigger)			
<i>Orsotriaena medus</i> (Fb.)	Common	Sri Lanka, S. India	Grasses
<b>Melanitis</b> (Evening browns)			
<i>Melanitis leda</i> (Drury)	Very common	All India, Sri Lanka	Grasses
<i>M. phedima</i> Cramer	Common	Sri Lanka, S. India	Grasses
<i>M. zitenius</i> (Herbst.)	Rare	S. India	Bamboos
<b>Parantirrhoea</b> (The Travancore evening brown)			
<i>Parantirrhoea marshalli</i> Wood Mason	Extremely local	Travancore, Coorg	<i>Ochlandra rheedii</i>
<b>Elymnias</b> (The Palmflies)			
<i>E. hypermnestra</i> Lin.	Common	Sri Lanka, Peninsular India	Palms
<b>AMATHUSIDAE</b>			
<b>Amathusia</b> (The Palmking)			
<i>Amathusia phidippus</i> (Johanssen)	Very rare	Travancore	Palms
<b>Discophora</b> (The Duffers)			
<i>Discophora sondaica tullia</i> (Cramer)	Very rare	N. Kanara	Bamboos
<i>D. lepida</i> (Moore)	Rare	Sri Lanka, S. India	Bamboos
<b>NYMPHALIDAE</b>			
<b>Charaxes</b> (The Rajahs)			
<i>C. bernardus imna</i> (Cramer)	Common	Peninsular India, W. Ghats, Nilgiris, Wynad	<i>Saccopetalum tomentosum</i> , <i>Tamarindus indica</i>
<i>C. solon solon</i> Fb.	Rare	Sri Lanka, India	<i>Tamarindus indica</i> , <i>Caesalpinia spicata</i>
<b>Polyura</b> (The Nawabs)			
<i>P. schreberi</i> (Godart)	Very rare	W. Ghats, Coorg, Travancore	<i>Caesalpinia spicata</i> , <i>Rourea minor</i>
<i>P. athama athama</i> S (Drury)	Common	Sri Lanka, Peninsular India	<i>Delonix regia</i> , <i>Caesalpinia</i> spp., <i>Albizia</i> spp., <i>Adenanthera pavonina</i>
<b>Rohana</b> (The Emperors)			

<i>Rohana parisatis</i> Fruh.	Common	Sri Lanka, W. Ghats, Nilgiris, Palni hills of Travancore Ghats, Nilgiris, Palnis	<i>Celtis tetrandra</i>
<b>Euthalia</b> (The Barons)			
<i>Tanaecia lepidea</i> (Butler)	Very rare	S. India	<i>Careya arborea</i> , <i>Melastoma malabathricum</i>
<i>E. telchinia</i> (Menetrieis)	Very rare	Coorg	
<i>E. alonthea meridionalis</i> Fruh.	Common	Sri Lanka, India	<i>Mangifera indica</i> , <i>Anacardium occidentale</i> , <i>Bryonia</i> spp., <i>Scurrula parasitica</i> , <i>Streblus asper</i>
<i>E. lubentina</i> (Cramer)	Rare	Sri Lanka, Peninsular, India	<i>Dendrophthoe falcata</i>
<i>E. evelina</i> (Stoll)	Rare	Sri Lanka, S. India	<i>Diospyros candolleana</i> , <i>D. melanoylon</i> , <i>Anacardium occidentale</i>
<b>Parthenos</b> (The Clipper)			
<i>Parthenos sylvia</i> Cramer	Common	Sri Lanka, W. Ghats, Nilgiris	<i>Modecca</i> sp., <i>Tinosperma cordifolia</i>
<b>Moduza</b> (The Admirals)			
<i>Moduza procris</i> (Cramer)	Common	Sri Lanka, Peninsular India	<i>Cinchona</i> , <i>Ochreinauclea missionis</i> , <i>Mussaenda frondosa</i> , <i>Wendlandia exserta</i> , <i>W. notoniana</i> , <i>Mitragyna parvifolia</i> , <i>Neolamarckia cadamba</i>
<b>Athyma</b> (The Sergeants)			
<i>Athyma nefte</i> Cramer	Common	S. India	<i>Glochidion</i> spp.
<i>A. selenophora</i> (Kollar)	Generally scarce	W. Ghats, Coorg	<i>Haldinia cordifolia</i>
<i>A. ranga</i> (Moore)	Rare	W. ghats, Nilgiris, Mysore, Palni, Wynad	<i>Olea dioica</i> , <i>Chionanthus</i>
<i>A. perius</i> (Linneaus)	Generally rare in India	Hills of Peninsular India	<i>Glochidion velutinum</i> , <i>G. lanceolatum</i>
<b>Neptis</b> (The Sailors)			
<i>Neptis columella</i>	Rare	W. Ghats, Coorg, Nilgiris, Palnis	<i>Dalbergia</i> sp.
<i>N. viraja kanara</i> Evans	Rare	India, Burma, Thailand, Andamans	
<i>N. nata hampsoni</i> Moore	Rare	India, Oriental region	
<i>N. clinia kallaura</i> Moore	Common	S. India, Nepal, Southern China	
<i>N. jumbah</i>	Common	Sri Lanka, S. India	<i>Xylia xylocarpa</i> , <i>Pongamia glabra</i> , <i>Dalbergia</i> sp., <i>Caesalpinia spicata</i> , <i>Bombax ceiba</i> , <i>Thespesia populnea</i> , <i>Hibiscus</i> sp., <i>Grewia</i> sp., <i>Elaeocarpus</i> sp., <i>Zizyphus</i> sp.
<i>N. hylas</i>	Very common	India, Sri Lanka	<i>Paracalya</i> sp., <i>Flemingia</i> sp., <i>Mucuna</i> sp., <i>Xylia</i> sp., <i>Caravalia mollis</i> , <i>Vigna unguicula</i> , <i>Grewia</i> sp., <i>Triumfetta</i> sp., <i>Corchorus</i> sp., <i>Bombax</i> sp., <i>Helicteres</i> sp., <i>Nothopodytis nimmoniana</i>
<i>N. soma</i>	Rare	W. Ghats, Hills of S. India	
<b>Pantoporia</b> (The Sergeant)			
<i>Neptis nata</i>	Rare	S. India	
<i>P. hordonia stoll</i>	Common	S. India	
<b>Cyrestis</b> (The Map butterflies)			

<i>Cyrestis thyodamas</i> Boisduval	Common	W. Ghats, Coorg, Wynad, Nilgiris, Palani	<i>Ficus religiosa</i> , <i>F. benghalensis</i> , <i>F. racemosa</i>
<b>Hypolimnas</b> (The Eggflies)			
<i>Hypolimnas bolina</i> (Lin.)	Common	India, Sri Lanka	<i>Laportea interrupta</i> ; <i>Elatostemma cunneatum</i> , <i>Portulaca oleracea</i>
<i>H. misippus</i> (Lin.)	Common	India, Sri Lanka	<i>Portulaca oleracea</i> , <i>Abutilon</i> sp.; <i>Hibiscus</i> sp.
<b>Doleschallia</b> (The Autumn leaf)			
<i>Doleschallia bisaltide</i> (Cramer)	Rare	Sri Lanka, W. Ghats, Nilgiris	<i>Acanthaceae</i>
<b>Kallima</b> (The Oakleaf butterflies)			
<i>Kallima philarchus</i> (Westwood)	Common	Sri Lanka, S. India	<i>Strobilanthes</i> spp.
<i>K. horsfieldi</i> Kollar	Rare	W. Ghats	
<b>Junonia</b> The Pansies)			
<i>Junonia hierta</i> (Fb.)	Very common	Sri Lanka, India	<i>Hygrophila auriculata</i> , <i>Barleria</i> sp.
<i>J. orithyia</i> (Lin.)	Very common	Sri Lanka, India	<i>Justicia procumbens</i> , <i>J. micrantha</i> , <i>Lepidagathis prostrata</i>
<i>J. lemonias</i> (Lin.)	Common	Sri Lanka, India	<i>Nelsonia campestris</i> , <i>Hygrophila auriculata</i> , <i>Corchorus</i> sp., <i>Sida rhombifolia</i>
<i>J. almana</i> (Lin.)	Common	Sri Lanka, India	<i>Hygrophila auriculata</i> , <i>Barleria</i> , <i>Hygrophila</i> , <i>Osbeckia</i> sp.
<i>J. atlites</i> (Johanssen)	Locally common	Sri Lanka, wetter regions of Peninsular India	<i>Hygrophila auriculata</i> , <i>Barleria</i>
<i>J. iphita</i> (Cramer)	Common	Sri Lanka, South and Central India	<i>Strobilanthes callosus</i> , <i>Justicia micrantha</i> , <i>H. aurioclata</i>
<b>Vanessa</b> (The Vanessas)			
<i>Vanessa cardui</i> (Lin.)	Common	Sri Lanka, India	<i>Zornia gibbosa</i> , <i>Artemisia</i> sp. <i>Blumea</i> sp. <i>Debregeasia</i> sp.
<i>V. indica</i> (Herbst)	Common	Sri Lanka, Hills of S. India, Coorg	<i>Girardinia heterophylla</i>
<b>Kaniska</b>			
<i>Kaniska canace</i> (Johanssen)	Common	Sri Lanka, Hills of S. India, Coorg	<i>Smilax</i> sp., Wild Yams
<b>Argyreus</b> (The Fritillaries)			
<i>Argyreus hyperbius</i> (Johanssen)	Common	Sri Lanka, Nilgiris, plains and hills of Cochin, Travancore	<i>Viola</i> sp.
<b>Phalanta</b> (The Leopards)			
<i>Phalanta phalantha</i> (Drury)	Common	India, Sri Lanka	<i>Flacourtia indica</i> , <i>F. montana</i> , <i>Aberia gardneri</i> , <i>Smilax</i> sp., <i>Salix</i> sp.
<i>P. alcippe</i> (Cramer)	Common	Sri Lanka, W. Ghats	<i>Alsodeia zeilanica</i>
<b>Cupha</b> (The Rustic)			
<i>Cupha erymanthis</i> (Drury)	Common	Sri Lanka, S. India	<i>Flacourtia indica</i> , <i>F. montana</i>
<b>Vindula</b> (The Cruiser)			
<i>Vindula erota</i> (Fb.)	Common	Sri Lanka, W. Ghats, Palnis, Nilgiris	<i>Adenia palmata</i>
<b>Cirrochroa</b> (The Yeoman)			
<i>Cirrochroa thais</i> (Fb.)	Common	Sri Lanka, W. Ghats, Coorg, Wynad, Nilgiris, Palnis	<i>Hydnocarpus pentandra</i>

<b>Cethosia</b> (The Lacewings)			
<i>Cethosia nietneri</i> Felder	Common	Sri Lanka, W. Ghats	<i>Adenia palmata</i>
<b>Ariadne</b> (The Castors)			
<i>Ariadne ariadne</i> (Johanssen)	Common	Sri Lanka, India	<i>Tragia involurata</i> , <i>T. cannabina</i> , <i>Ricinus communis</i>
<i>A. merione</i> (Cramer)	Common	Sri Lanka, India	<i>Tragia involucrata</i> , <i>T. cannabina</i> , <i>Ricinus communis</i>
<b>ACRAEIDAE</b>			
<b>Acraea</b> (The Tawny coster)			
<i>Acraea violae</i> (Fb.)	Common	Sri Lanka, India	Cultivated passion flower, <i>Adenia palmata</i> , <i>Passiflora foetida</i> , <i>Hibiscus cannabinus</i> , <i>Cucurbits</i>
<b>LIBYTHEIDAE</b>			
<b>Libythea</b> (The Beaks)			
<i>Libythea myrrha</i> Godart	Common	Sri Lanka, S. India	<i>Celtis tetrandra</i>
<i>L. lepita</i> Moore	Rare	Sri Lanka, S. India	<i>Celtis australis</i>
<b>RIODINIDAE</b>			
<b>Abisara</b> (The Judies)			
<i>Abisara echerius</i> (Stoll)	Common	Sri Lanka, Peninsular India	<i>Embelia robusta</i> , <i>Ardisia</i> sp.
<b>LYCAENIDAE</b>			
<b>Spalgis</b> (The Apefly)			
<i>Spalgis epius</i> (Westwood)	Common	Sri Lanka, W. Ghats, Coorg, Bangalore	Carnivorous on <i>Coccidae</i>
<b>Talicauda</b> (The Red pierrot)			
<i>Talicauda nyseus</i> (Guerin)	Locally abundant	Sri Lanka, S. India	<i>Bryophyllum calycinum</i> , <i>Kalanchoe laciniata</i>
<b>Castalius</b> (The Pierrots)			
<i>Castalius rosimon</i> (Fabricius)	Common	Sri Lanka, India	<i>Zizyphus rugosa</i> , <i>Z. jujuba</i>
<b>Caleta</b>			
<i>C. caleta</i> (Hewitson)	Common	Sri Lanka, Peninsular India	<i>Zizyphus rugosa</i>
<b>Discolampa</b>			
<i>Discolampa ethion</i> (Doubleday and Hewitson)	Common	Sri Lanka, S. India	<i>Zizyphus oenoplia</i> , <i>Z. jujuba</i>
<b>Tarucus</b> (The Blue pierrots)			
<i>Tarucus ananda</i> (de Niceville)	Rare	Nilgiris, Palnis, Annamalai Hills	<i>Zizyphus xylopyrus</i> , <i>Loranthus</i> spp.
<i>T. nara</i>	Common	Sri Lanka, S. India	<i>Zizyphus jujuba</i>
<b>Syntarucus</b> (The Zebra-blue)			
<i>Syntarucus plinius</i> Fb.	Common	Sri Lanka, India	<i>Albizia lebbek</i> , <i>Indigofera</i> spp., <i>Sesbania aculeata</i>
<b>Azarus</b> (The Babul blues)			
<i>Azarus ubaldus</i> (Cramer)	Common	India, Sri Lanka	<i>Acacia arabica</i> , <i>A. leucophloea</i>
<i>A. uranus</i> Butler	Not common	S. India	<i>Acacia senegal</i>
<i>A. jesous</i> (Guerin)	Common	Sri Lanka, India	<i>Acacia farnesiana</i> , <i>A. leucophloea</i>
<b>Neopithecopis</b> (The Quaker)			
<i>Neopithecopis zalmora</i> (Butler)	Common	Sri Lanka, S. India	<i>Glycosmis mouritiana</i>

<b>Megisba</b> (The Malayan)			
<i>M. malaya thwaitesi</i>		S. India	
<b>Acytolepis</b> (The Hedge blues)			
<i>Acytolepis puspa</i>	Common	Sri Lanka, Peninsular India	<i>Cylista scariosa</i> , <i>Xylia dolabiformis</i> , <i>Hiptage madablate</i> , <i>Schleichera trjuga</i>
<i>A. lilacea</i>	Rare	Nilgiris, Palnis, Coorg, Sri Lanka	
<i>Udara akasa</i>	Locally common	Sri Lanka and hills of S. India	
<i>Celatoxia albidisca</i>	Fairly common	Hills of S. India	
<i>Celastrina lavendularis</i>	Common	S. India, Sri Lanka	
<b>Chilades</b> (The Lime blue)			
<i>Chilades lajus</i> (Cramer)	Common	Sri Lanka, India	<i>Limonia acidissima</i> , young shoots of orange
<i>C. pandava</i> (Horsfield)	Locally common	India, Sri Lanka	<i>Xylia xylocarpa</i> , <i>Cycas revoluta</i> , Lime etc.
<i>C. parhasius</i>	Common	Indian sub continent	
<b>Freyeria</b>			
<i>Freyeria trochylus</i>	Common	Sri Lanka, India	<i>Lotus corniculatus</i> , <i>Indigo peapoda</i> , <i>Vetches</i> sp., <i>Rhynchosia minima</i> , <i>Heliotropium strigosum</i>
<b>Pseudozizeeria</b>			
<i>P. maha</i>	Common	S. India	<i>Oxalis corniculata</i> , <i>Tephrosia pauciflora</i> , <i>Nelsonia</i> sp., <i>strobilanthes</i> sp.
<i>P. lysimon</i>	Common	Sri Lanka, India	<i>Zornia gibbosa</i> , <i>Amaranthus viridis</i>
<i>Zizula hylax hylax</i> Fb.	Common	Sri Lanka, India	<i>Nelsonia campestris</i> , <i>Lantana camara</i>
<b>Zizina</b>			
<i>Zizina</i>	Common	India, Sri Lanka	<i>Zornia gibbosa</i> , <i>Sesbania aculeata</i> , <i>Alysicarpus vaginalis</i>
<b>Euchrysops</b> (The Plains cupids)			
<i>Euchrysops cnejus</i> (Fabricius)	Common	India, Sri Lanka	<i>Ougenia dalbergioides</i> , <i>Phaseolus trilobus</i> , <i>Paracalyx scariosa</i> , <i>Butea frondosa</i> , <i>Acacia</i> , Peas, Beans, Grams, and <i>Vigna unguiculata</i>
<i>E. contracta</i> (Butler)	Common	Sri Lanka, S. India	
<b>Catachrysops</b>			
<i>Catachrysops strabo</i> (Fb.)	Common	India, Sri Lanka	<i>Ougenia dalbergioides</i> , <i>Paracalyx scariosa</i> , <i>Schleichera oleosa</i>
<b>Lampides</b> (The Pea blue)			
<i>Lampides boeticus</i> (Lin.)	Very common	India, Sri Lanka	<i>Butea frondosa</i> , <i>Crotalaria capensis</i> , <i>Melotus</i> sp., beans, peas
<b>Anthene</b>			
<i>Anthene emolus</i> (Godart)	Rare	S. India	<i>Combretum latifolium</i> , <i>Terminalia paniculata</i> , <i>Trichilia connaroides</i> , <i>Saraca indica</i> , <i>Cassia fistula</i> , <i>Litchi chinensis</i>
<i>A. lycaenina</i> Felder	Rare	S. India, Sri Lanka	<i>Caesalpinia spicata</i> , <i>Acacia pennata</i> , <i>Buchanania lanzan</i> , <i>B. angustifolia</i>
<b>Jamides</b> (The Ceruleans)			

<i>J. celeno</i>	Common	India, Sri Lanka	<i>Pongamia glabra</i> , <i>Abrus precatorius</i> , <i>Saraca indica</i> , <i>Butea frondosa</i> , <i>Trichilia connaroides</i>
<i>J. alecto</i>	Common	Sri Lanka, S. India	<i>Kaempferia pandurata</i> , <i>Elettaria cardamoum</i>
<i>J. bochus bochus</i> Cramer	Common	Sri Lanka, S. India	<i>Pongamia glabra</i> , <i>Xylia xylocarpa</i> , <i>Crotalaria</i> sp., <i>Butea frondosa</i> ; <i>Vigna unguiculata</i> , <i>Tephrosia candida</i>
<b>Nacaduba (The Line blues)</b>			
<i>Nacaduba pactolus</i>	Rare	Coorg, Nilgiris, Sri Lanka	
<i>N. hermus</i>	Common	Sri Lanka, India	
<i>N. kurava</i>	Rare	Sri Lanka, India	<i>Embelica tsjeriam-cottam</i> , <i>Ardisia solanacea</i> , <i>Waltheria indica</i>
<i>N. beroe</i>	Common	Sri Lanka, India	<i>Caesalpinia spicata</i>
<i>N. calauria</i>	Rare	Sri Lanka, S. India	
<i>N. berenice ormistoni</i> Toxopeus	Common	Sri Lanka	
<b>Prosotas</b>			
<i>P. nora</i>	Common	India, Sri Lanka	<i>Acacia caesia</i> , <i>Combretaceae</i> , <i>Myrtaceae</i> , <i>Sapindaceae</i>
<i>P. dubiosa indica</i> Evans	Common	Oriental region	
<i>P. noreia</i>	Very rare	Sri Lanka, S. India	
<b>Petrelaena</b>			
<i>Petrelaea dana</i>	Common	Sri Lanka, S. India	
<b>Ionolyce</b>			
<i>Ionolyce helicon</i>	Rare	Sri Lanka, India	
<b>Curetis (The Sunbeams)</b>			
<i>Curetis thetis</i> (Drury)	Common	Sri Lanka, Peninsular India	<i>Pongamia glabra</i> , <i>Derris scandens</i> , <i>Xylia xylocarpa</i> , <i>Abrus precatorius</i> , <i>Trichilia connaroides</i>
<i>C. dentata</i> (Moore)	Common	S. India	
<i>C. siva</i>	Rare	Sri Lanka, Coorg, Mysore, Nilgiris, Travancore	<i>Ougenia dalbergioides</i>
<b>Iraota (The Silverstreak blues)</b>			
<i>Irota timoleon</i> (Stoll)	Common	S. India	<i>Punica granatum</i>
<b>Amblypodia (The Leaf blue)</b>			
<i>Amblypodia anita</i> Hewitson	Common	Sri Lanka, Peninsular India	<i>Olea</i> sp.
<b>Thaduka (The Many-tailed Oak blue)</b>			
<i>Thaduka multicaudata</i> (Moore)	Rare	Kanara, Coorg, Malabar, W. slopes of Nilgiris	<i>Trewia nudiflora</i>
<b>Arhopala (The Oak blues)</b>			
<i>Arhopala bazaloides</i>	Very rare	W. Ghats	
<i>A. amantes</i>	Locally common	Sri Lanka, S. India	<i>Terminalia crenulata</i> , <i>T. paniculata</i> , <i>T. catappa</i> , <i>Lagerstroemia microcarpa</i> , <i>L. hirsuta</i> , <i>Hopea jucunda</i>
<i>A. pseudocentaurus</i>	Common	Sri Lanka, S. India	<i>Terminalia crenulata</i> , <i>T. paniculata</i> , <i>Lagerstroemia microcarpa</i> , <i>Xylia xylocarpa</i>
<i>A. canaraica</i>	Very rare	Travancore, Nilgiris, Coorg, Kanara	

<i>A. abseus</i>	Very rare	Sri Lanka, Kanara, India, <i>Shorea robusta</i>	
<b>Surendra</b> (The Acacia blues)			
<i>Surendra quercetorum</i> Moore	Fairly common	Sri Lanka, W. Ghats, Palnis, Nilgiris	<i>Acacia pennata</i> , <i>A. caesia</i>
<b>Zinaspa</b>			
<i>Z. todara</i> Moore	Rare	Nilgiris, Kannara	
<b>Loxura</b> (The Yamfly)			
<i>Loxura atymnus</i> (Cramer)	Common	Sri Lanka, Nilgiris, Palnis, Coorg, Shevaroyis, Travancore Cochin Hills	<i>Dioscorea pentaphylla</i> , <i>Smilax</i> sp.
<b>Spindasis</b> (The Silverlines)			
<i>Spindasis vulcanus</i>	Common	India, Sri Lanka	
<i>S. lilacinus</i> Moore	Rare	Mysore, Bangalore	<i>Canthium parviflorum</i> , <i>Zizyphus rugosa</i> , <i>Z. jujuba</i>
<i>S. schistacea</i>	Common	S. India, Sri Lanka	
<i>S. ictis</i>	Locally common	Sri Lanka, India	
<i>S. elima</i>	Rare	India, Sri Lanka	
<i>S. abnormis</i> Moore	Rare	South India	
<i>S. lohita</i>	Common	Sri Lanka, W. Ghats, Nilgiris, Palanis, Bangalore	<i>Terminalia paniculata</i> , <i>Xylia</i> , <i>Dioscorea pentaphylla</i> , <i>Convolvulaceae</i>
<b>Zesius</b> (The Redspot)			
<i>Zesius chrysomallus</i> Hubner	Common	Sri Lanka, Peninsular India	<i>Terminalia paniculata</i> , <i>Pterocarpus marsupium</i> , <i>Anacardium occidentale</i> , <i>Psidium guava</i> , <i>Loranthaceae</i>
<b>Ancema</b>			
<i>Ancema blanka</i> (de Niceville)	Rare	S. India	<i>Viscum angulatum</i> , <i>V. capitellatum</i>
<b>Pratapa</b> (The Tufted royals)			
<i>P. deva</i> (Moore)	Rare	S. India, Sri Lanka	<i>Scurrula parasitica</i> , <i>Taxillus tomentosus</i> , <i>Dendrophthoe falcata</i>
<b>Creon</b>			
<i>C. cleobis</i> (Godart)	Rare	S. India, W. Ghats, Coorg, Nilgiris	<i>Loranthus elasticus</i> , <i>Viscum capitellatum</i>
<b>Tajuria</b> (The Royals)			
<i>Tajuria melastigma</i> de Niceville	Rare to very rare	N. Kanara, Nilgiris	
<i>T. jehana</i> Moore	Common	Sri Lanka, S. India	
<i>T. maculata</i> Hewitson	Common	S. India	
<i>T. cippus</i> (Fb.)	Fairly common	India, Sri Lanka	<i>Dendrophthoe falcata</i> , <i>L. elasticus</i> , <i>Helixanthera wallichiana</i>
<b>Eliotia</b> (The Mandarin blues)			
<i>Rachana jalindra</i> (Horsfield)	Very rare	S. India, (N. Kanara, Coorg, Nilgiris)	<i>Loranthus elasticus</i>
<b>Cheritra</b> (The Common imperial)			
<i>Cheritra freja</i> (Fb.)	Common	Sri Lanka, W. Ghats	<i>Xylia xylocarpa</i> , <i>Saraca indica</i> , <i>Cinnamomum</i> sp., <i>Ixora</i> sp.
<b>Rathinda</b> (The Monkey puzzle)			



<i>Rathinda amor</i> (Fb.)	Common	Sri Lanka, S. India	<i>Hopea</i> sp., <i>Loranthus</i> , <i>Croton</i> , <i>Blachia</i> , <i>Schleicheria</i> sp., <i>Litchi chinensis</i> , <i>Careya</i> sp., <i>Eugenica zeylanica</i>
<b>Horaga</b> (The Onyxes)			
<i>Horaga onyx</i> (Moore)		Scarce in S. India, Sri Lanka	<i>Coriaria nepalensis</i>
<i>H. viola</i> Moore	Scarce	S. India	<i>Coriaria nepalensis</i>
<b>Catapaecilma</b> (The Tinsels)			
<i>Catapaecilma elegans</i> (Druce)	Common	Sri Lanka, S. India	<i>Terminalia paniculata</i>
<b>Hypolycaena</b> (The Tits)			
<i>Hypolycaena nilgirica</i> Moore	Very rare	Sri Lanka, Nilgiris, Palnis	
<i>H. othona</i> (Hewitson)	Rare	W. Ghats	Orchids, <i>Cottonia peduncularis</i> , <i>Aerides crispum</i> , <i>Rhynchostylis retusa</i>
<b>Zeltus</b> (The fluffy tit)			
<i>Zeltus amasa amasa</i> Hewitson	Not common	W. Ghats	
<b>Deudorix</b> (The Cornelians)			
<i>Deudorix epjarbas</i> (Moore)	Rare	Sri Lanka, India	Pomegranate fruits, <i>Sapindus trifoliatus</i> , <i>Aesculus indicus</i> , <i>Connarus</i> sp.
<b>Virachola</b>			
<i>Virachola isocrates</i> (Fb.)	Common	Sri Lanka, S. India	Fruits of <i>Catunaragam spinosa</i> , <i>Eriobotria japonica</i> , <i>Psidium guava</i> , <i>Tamarindus indica</i> , <i>Strychnos nux-vomica</i> , <i>Limonia acidissima</i> , <i>Punica granatum</i>
<i>V. perse</i> (Hewitson)	Common	Sri Lanka, S. India	Fruits of <i>Catunaragam spinosa</i>
<b>Rapala</b> (The Flashes)			
<i>Rapala varuna</i>	Rare	Sri Lanka, S. India	Flowers of <i>Zizyphus xylopyrus</i> , <i>Z. rugosa</i> , <i>Quisqualis indica</i> , <i>Sapindus trifoliata</i>
<i>R. manea schistacea</i>	Common	Sri Lanka, India	Flowers of <i>Spiraea sorbifolia</i> , <i>Antidesma acidum</i> , <i>Zizyphus</i> , <i>Acacia pennata</i> , <i>A. caesia</i> , <i>Quisqualis indica</i> , Tea blossoms
<i>R. lankana</i>	Very rare	Sri Lanka, W. Ghats, Nilgiris	
<i>R. iarbus</i> Fb.	Common	Sri Lanka, S. India	Flowers of <i>Ougenia dalbergioides</i> , <i>Zizyphus</i> sp.
<b>Bindahara</b> (The Plane)			
<i>Bindahara phocides</i> (Moore)	Rare	S. India, Sri Lanka	Inside fruits of <i>Salacia macrosparma</i> and <i>S. reticulata</i>
<b>PAPILIONIDAE</b>			
<b>Troides</b> (The Birdwings)			
<i>Troides minos</i> Cramer	Common	S. India	<i>Aristolochia indica</i> , <i>Apama siliquosa</i>
<b>Pachliopta</b> (The Red bodied swallow tails)			
<i>Pachliopta pandiyana</i> Moore	Locally common	W. Ghats	<i>Apama siliquosa</i>
<i>P. hector</i> (Lin.)	Common	S. India	<i>Aristolochia indica</i>
<i>P. aristolochiae</i> (Fb.)	Very common	India, Sri Lanka	<i>Aristolochia indica</i>
<b>Chilasa</b> (The Mimes)			
<i>Chilasa clytia</i> (Lin.)	Common	S. India, Sri Lanka	<i>Alseodaphne semicarpifolia</i> , <i>Cinnamomum zeylanicum</i> , <i>Litsea tomentosa</i> , <i>L. glutinosa</i>

<b>Papilio</b> (The Black-bodied swallowtails)			
<i>Papilio polymnestor</i> Cramer	Common	Sri Lanka, S. India	<i>Citrus decumana</i> , <i>Glycomis mouritiana</i> , <i>Atalantia</i> spp.
<i>P. paris</i> Lin.	Common	W. Ghats, Oriental region	<i>Evodia</i> sp.
<i>P. crino</i> Fb.	Rare	Sri Lanka, S. India	<i>Chloroxylon swietenia</i>
<i>P. buddha</i> Westwood	Local and generally rare	W. Ghats	<i>Zanthoxylum rhetsa</i>
<i>P. dravidarum</i> Wood-Mason	Rare	W. Ghats	<i>Glycosmis mouritiana</i>
<i>P. helenus</i> Lin.	Common	Sri Lanka, W. Ghats	Cultivated citrus, <i>Zanthoxylum rhetsa</i> , <i>Glycosmis mouritiana</i>
<i>P. polytes</i> Lin.	Very common	Sri Lanka, India	Cultivated citrus <i>Glycosmis mouritiana</i> , <i>Citrus medica</i> , <i>Aegle marmelos</i> , <i>Zanthoxylum rhetsa</i> , <i>Murraya koenigii</i>
<i>P. liomedon</i> Moore	Rare	W. Ghats	<i>Acronychia pedunculata</i>
<i>P. demoleus</i> Lin.	Very common	Sri Lanka, India	Cultivated citrus, <i>Ruta graveoleus</i> <i>Aegle mormelos</i> , <i>Glycosmis mouritiana</i>
<b>Pathysa</b>			
<i>Pathysa nomius</i> (Esper)	Common	Sri Lanka, Peninsular India	<i>Saccopetalum tomentosum</i> , <i>Polyalthia longifolia</i>
<i>P. antiphates</i> (Cramer)	Rare	Sri Lanka, S. India	<i>Desmos lawii</i>
<b>Graphium</b> (The kite swallowtails)			
<i>Graphium sarpedon</i> (Lin.)	Common	Sri Lanka, S. India	<i>Persia oderatissima</i> , <i>P. micrantha</i> <i>Cinnamomum zeylanicum</i> , <i>Camphora officinalis</i> , <i>Litsea chinensis</i> , <i>L. salicifolia</i> , <i>Alesodaphne semicarpitulia</i> , <i>Litsea salicifolia</i>
<i>G. doson</i> (C. & R. Felder)	Common	Sri Lanka, S. India	<i>Saccopetalum tomentosum</i> , <i>Desmos lawii</i> , <i>Polyalthia longifolia</i> , <i>Cinnamomum</i> sp.
<i>G. agamemnon</i> (Lin.)	Common	Sri Lanka, S. India	<i>Saccopetalum tomentosum</i> , <i>Anona muricata</i> , <i>A. squamosa</i> , <i>A. discolor</i> , <i>A. reticulata</i> , <i>Polyalthia longifolia</i> , <i>Saccopetalum</i> sp. <i>Gautheria</i> sp.
<b>PIERIDAE</b>			
<b>Leptosia</b> (The Psyche)			
<i>Leptosia nina</i> (Fb.)	Common	India, Sri Lanka	<i>Capparis rheedii</i> , <i>Crataeva magna</i>
<b>Delias</b> (The Jezebels)			
<i>Delias eucharis</i> (Drury)	Common	Sri Lanka, India	<i>Loranthus elasticus</i> , <i>Scurrula parasitica</i> , <i>Dendrophthoe falcata</i>
<b>Cepora</b> (The Gulls)			
<i>Cepora nerissa</i> (Fb.)	Common	Sri Lanka, India	<i>Capparis decidua</i> , <i>C. sepiaris</i> , <i>C. rheedii</i> , <i>C. zeylanica</i>
<i>C. nadina</i> (Lucas)	Common	Sri Lanka, W. Ghats	<i>Capparis roxburghii</i> , <i>C. moonii</i> , <i>C. rheedii</i>
<b>Prioneris</b> (The Saw-tooths)			
<i>Prioneris sita</i> (C. & R. Felder)	Generally	Sri Lanka, W. Ghats	<i>Capparis zeylanica</i>
<b>Anaphaeis</b>			
<i>Anaphaeis aurota</i> (Fb.)	Common	Sri Lanka, India	<i>Capparis decidua</i> , <i>C. sepiaria</i> , <i>C. rheedii</i> , <i>Cadaba indica</i> , <i>Macrua arenaria</i>
<b>Appias</b> (The Puffins and Albatrosses)			
<i>Appias indra</i> (Moore)	Rare	Sri Lanka, S. India	<i>Drypetes</i> sp.

<i>A. libythea</i> (Fb.)	Common	Sri Lanka, S. India	<i>Crataeva magna</i> , <i>Capparis sepiaria</i> , <i>C. roxburghii</i> , <i>C. zeylanica</i>
<i>A. lyncida</i> (Cramer)	Scarce and local	S. India, Sri Lanka	<i>Crataeva magna</i> , <i>Capparis roxburghii</i>
<i>A. albina</i> (Boisduval)	Common	Sri Lanka, S. India	
<i>A. wardi</i> (Moore)	Rare	S. India	<i>Drypetes venusta</i>
<b>Artogeia</b>			
<i>Artogeia canidia</i> (Sparrman)	Very common	Nilgiris, Palni hills, Cochin and Travancore	Cruciferae
<b>Ixias (The Indian orange tips)</b>			
<i>Ixias marianne</i> (Cramer)	Common	Sri Lanka, Peninsular India	<i>Capparis sepiaria</i> , <i>C. divaricata</i> , <i>C. decidua</i> , <i>C. grandis</i>
<i>I. pyrine</i> (Lin.)	Common	Sri Lanka, Peninsular India	<i>Capparis sepiaria</i>
<i>I. pyrene sesia</i> Fb.	Common	Sri Lanka, India, Hong Kong, Malaysia	
<b>Colotis (The Arabs and Small orange tips)</b>			
<i>Colotis amata</i> Fb.	Common	Sri Lanka, India	<i>Salvadora persica</i> , <i>S. oleoides</i> , <i>Azima tetraacantha</i>
<i>C. fausta</i> (Olivier)	Common	Peninsular India	<i>Maerua oblongifolia</i>
<i>C. etrida</i> (Boisduval)	Common	Sri Lanka, Palnis	<i>Cadaba indica</i>
<i>C. eucharis</i> (Fb.)	Common	Sri Lanka, Peninsular India	<i>Cadaba indica</i>
<i>C. danae</i> (Fb.)	Common	Sri Lanka, Peninsular India	<i>Cadaba indica</i> , <i>Maerua oblongifolia</i> , <i>Capparis sepiaria</i> , <i>C. divaricata</i>
<b>Hebomoia (The Great orange tip)</b>			
<i>Hebomoia glaucippe</i> (Lin.)	Common	Peninsular India, Sri Lanka	<i>Crataeva magna</i> , <i>Capparis mooni</i>
<b>Pareronia (The Wanderers)</b>			
<i>Pareronia ceylanica</i> (C. & R. Felder)	Common	Sri Lanka, Mysore, W.Ghats, Nilgiris	
<i>P. valeria</i> (Cramer)	Common	India	
<b>Catopsilia (The Emigrants)</b>			
<i>C. pomona</i> (Fb.)	Very common	Sri Lanka, India	<i>Cassia fistula</i> , <i>C. siamea</i> , <i>C. tora</i> , <i>Butea frondosa</i> , <i>Bauhinia racemosa</i>
<i>C. pyranthe</i> (Lin.)	Very common	Sri Lanka, India	<i>Cassia tora</i> , <i>C. auriculata</i> , <i>C. occidentalis</i>
<b>Eurema (The Grass yellows)</b>			
<i>Eurema brigitta</i> (Cramer)	Very common	Sri Lanka, India	<i>Cassia kleinii</i>
<i>E. laeta</i> Boisduval	Very common	India, Sri Lanka	Leguminosae
<i>Eurema hecabe</i> (Lin.)	Very common	India, Sri Lanka	<i>Cassia tora</i> , <i>C. fistula</i> , <i>Caesalpinia spicata</i> , <i>Pithecolobium dulce</i> , <i>Sesbania aculeata</i> , <i>Caesalpinia</i> spp., <i>Albizia</i> spp.
<i>E. blanda</i> Boisduval	Common	Sri Lanka, Peninsular India	<i>Caesalpinia spicata</i> , <i>Delonix regia</i> , <i>Cassia</i> spp., <i>Albizia</i> spp.
<i>E. andersoni</i> Moore	Rare	Sri Lanka, Peninsular India	
<b>Colias (The Clouded yellows)</b>			
<i>Colias nilagiriensis</i> Felder & Felder	Common	Hills of S. India	<i>Parochetus communis</i>

<b>HESPERIPDAE</b>			
<b>Celaenorrhinus (The Flats)</b>			
<i>Celaenorrhinus leucocera</i> (Kollar)	Common	S. India	<i>Eranthemum roseum</i> , <i>Ecbolium viride</i> , <i>Strobilanthes callosus</i> , <i>S. angustifrons</i>
<i>C. ruficornis</i> (Mabille)	Common	W. Ghats, Nilgiris, Palnis	<i>Strobilanthes callosus</i>
<i>C. ambareesa</i>	Common	S. India	<i>Strobilanthes callosus</i> , <i>Eranthemum roseus</i> , <i>E. purpurascens</i>
<b>Tagiades (The Snow flats)</b>			
<i>Tagiades gana silvia</i> Evans	Common	S. India	<i>Dioscorea oppositifolia</i>
<i>T. japetus</i> Mabille	Common	Sri Lanka, Nilgiris, Shevaroy's, Palnis	
<i>T. litigiosa</i> Moschler	Common	Sri Lanka, Peninsular India	<i>Dioscorea oppositifolia</i> , <i>Smilax</i>
<b>Coladenia (The Pied flats)</b>			
<i>Pseudocoladenia dan</i> (Fb.)	Common	S. India	<i>Achyranthes aspera</i>
<i>P. indrana indra</i> Evans	Rare	Sri Lanka, S. India, Nepal, Burma	
<b>Sarangesa (The small flats)</b>			
<i>Sarangesa dasahara</i> (Moore)	Common	Sri Lanka, S. India	<i>Asystasia</i> spp., <i>Blepharis asperina</i>
<i>S. purendra pandra</i> Evans	Common	Indian sub-continent	
<b>Odontoptilum (The Chestnut angle)</b>			
<i>Odontoptilum angulata</i> (Felder)	Common	Peninsular India	<i>Allophylus cobbe</i>
<b>Caprona (The Golden angle)</b>			
<i>Caprona ransonnetti</i> (Felder)	Common	S. India	<i>Helicteres isora</i>
<i>C. alida vespa</i> Evans	Rare	Sri Lanka, S. India, Nepal, South China and Hong Kong	
<b>Spiatia (The Indian skipper)</b>			
<i>Spiatia galba</i> (Fb.)	Common	India, Sri Lanka	<i>Waltheria indica</i> , <i>Sida rhombifolia</i> , <i>Hibiscus</i> sp.
<b>Hasora (The Common banded awl)</b>			
<i>Hasora chromus</i> Cramer	Common	India, Sri Lanka	<i>Pongamia glabra</i> , <i>Ricinus communis</i> , <i>Trichilia trifoliata</i>
<i>H. taminatus</i> (Hubner)	Common	W. Ghats, Coorg, Palnis, Nilgiris, Sri Lanka	<i>Derris scandens</i>
<i>H. badra</i> (Moore)	Common	Sri Lanka, W. Ghats	<i>Derris uliginosa</i>
<b>Ismene (The Orange awlet)</b>			
<i>Ismene jaina</i> (Moore)	Fairly	W. Ghats, Sri Lanka	<i>Combretum latifolium</i>
<b>Bibasis (The Orange tail awl)</b>			
<i>Bibasis sena</i> (Moore)	Rare	Sri Lanka, Coorg, Nilgiris	<i>Combretum latifolium</i> , <i>Hiptage madablota</i>
<b>Badamia (The Brown awl)</b>			
<i>Badamia exclamations</i> (Fb.)	Common	India, Sri Lanka	<i>Terminalia bellirica</i> , <i>Combretum ovalifolium</i> , <i>Chionanthes purpurea</i> , <i>Ficus</i> spp.
<b>Suastus (The Indian palm Bobs)</b>			
<i>Suastus gremius</i> (Fb.)	Common	India, Sri Lanka	Palms

<i>S. minuta bipunctus</i> Swinhoe	Common	Sri Lanka, S. India, Sikkim, Philippines and Java	
<b>Arnetta</b> (The Bobs)			
<i>Arnetta vindhiana</i> (Moore)	Common	South to Central India	
<i>A. mercara</i> Evans	Rare	W. Ghats	
<b>Matapa</b> (The Common red eye)			
<i>Matapa aria</i> (Moore)	Common	S. India	Bamboos
<b>Erionota</b> (The Palm red eye)			
<i>Erionota thra</i> (Hubner)	Very rare	S. India	<i>Musa sapientium</i>
<b>Taractrocera</b> (The Grass darts)			
<i>Taractrocera maevius</i> (Fb.)	Common	India, Sri Lanka	Grasses
<i>T. ceramas</i> (Hewitson)	Common	W. Ghats, Hills of S. India	Grasses
<b>Aeromachus</b> (The Scrub hoppers)			
<i>Aeromachus pygmaeus</i> (Fb.)	Common	Nilgiris, Wynad, Coorg (S. India)	Grasses
<b>Ampittia</b> (The Bush hoppers)			
<i>Ampittia dioscorides</i> (Fb.)	Fairly common	Sri Lanka, Peninsular India	Grasses, Paddy
<i>A. pygmaeus</i> Fb.	Common	S. India	
<b>Telicota</b> - (The Palm darts)			
<i>Telicota colon</i> Fb.	Common	Sri Lanka, India	Sugarcane
<i>T. ancilla</i> Moore	Common	Sri Lanka, India	Bamboos
<b>Oriens</b> (The Dartlets)			
<i>Oriens goloides</i> (Moore)	Common	S. India, Sri Lanka	Grasses
<i>O. concirna</i> (Elwes)	Rare	Shevaroys, Nilgiris, Palnis, Coorg	
<b>Potanthus</b> (The Darts)			
<i>Potanthus pseudomaesa</i> Moore	Abundant	Plains of India, Sri Lanka	Grasses
<i>P. confucius</i> Evans	Abundant	Sri Lanka, Hills of S. India	
<i>P. pallida</i> Evans		Sri Lanka, S. India, China and Yunan	
<i>P. palnia</i> Evans	Abundant	S. Indian hills	
<i>P. pava pava</i> Fruhstofer		S. India, Himachal Pradesh and Oriental region	
<b>Halpe</b>			
<i>Halpe homolea</i> (Hewitson)	Common in S. India	Travancore, S. India, Palni, Nilgiris, Wynad, Bangalore, Sri Lanka	Bamboos
<i>H. moore</i> Watson	Common	Nilgiris, Coorg	
<i>H. honorei</i> de Niceville	Rare	Palnis, Nilgiris, Trichinapally, Coorg	
<b>Baracos</b> (The Hedge hoppers)			
<i>Baracus vittatus</i> (Felder)	Common	Sri Lanka, Nilgiris, Palnis, Travancore Hills	
<i>B. hampsoni</i> Elwes	Common	W. Ghats	Grasses

<b>Baoris (The swifts)</b>			
<i>Baoris farri</i>	Common in S. India	W. Ghats	Bamboos
<b>Caltoris</b>			
<i>Caltoris canaraica</i>	Common	Coorg, Wynad, N. Kanara	Bamboos
<i>C. philippina</i>	Rare	Nilgiris, Coorg, N. Kanara, Sri Lanka	Bamboos
<i>C. kumara</i>	Common	W. Ghats, Sri Lanka	Bamboos
<b>Polytremis</b>			
<i>Polytremis lubricans</i>	Not common	Travancore, Nilgiris, Wynad, Coorg	
<i>P. lubricans lubricans</i> Herrich Schaffer		W. Ghats, Kumaon, China, Sula Islands	
<b>Pelopidas</b>			
<i>Pelopidas conjuncta</i>	Common	Sri Lanka, Peninsular India	Bamboos and grasses
<i>P. subochracea</i>	Rare	Sri Lanka, Coorg, Palnis, Nilgiris	Grasses
<i>P. mathias</i>	Very common	Sri Lanka, India	Grasses
<i>P. guttatus</i>	Very common	Sri Lanka, India	Grasses
<i>P. agna</i> Moore	Common	Oriental region	
<b>Borbo</b>			
<i>Borbo cinnara</i>	Common	Sri Lanka, India	Grasses
<i>B. bevant</i>	Locally common	S. India	
<b>Notocrypta (The Demons)</b>			
<i>Notocrypta paralysos</i> (Wood Mason)	Fairly common	Sri Lanka, W. Ghats	Scitaminae
<i>N. curvifascia</i> (Felder)	Common	Sri Lanka, W. Ghats, Mysore, Palni	<i>Kaempferia rotunda</i> , <i>Zingiber cossumnar</i> , <i>Curcuma decipiens</i>
<b>Udaspes (The Grass demon)</b>			
<i>Udaspes folus</i> (Cramer)	Common	Sri Lanka, S. India	<i>Curcuma decipiens</i> , Grasses
<b>Hyarotis (The Tree flitter)</b>			
<i>Hyarotis adrastus</i> (Cramer)	Rare	Sri Lanka, W. Ghats	<i>Phoenix aculis</i>
<i>H. microstictum coorga</i> Evans			
<b>Psolos (The Coon)</b>			
<i>Psolos fuligo</i> (Mabille)	Common	W. Ghats	<i>Phrynium</i>
<b>Iambrix (The Chestnut bob)</b>			
<i>Iambrix salsala</i> (Moore)	Common	Sri Lanka, S. India	Grasses